



Synthesis of Pent-aza Phenoxazines: A New Group of Anti-inflammatory Heterocycles. Their Effect and Other Analgesics on Pain

E. M. Odin^{1*}, P. K. Onojah¹, L. E. S. Akpanisi² and B. O. Akabueze¹

¹Department of Pure and Industrial Chemistry, Kogi State University, Nigeria.

²Department of Pure and Industrial Chemistry, University of Nigeria, Nigeria.

Authors' contributions

This work was carried out in collaboration between all authors. All authors read and approved the final manuscript.

Article Information

DOI: 10.9734/ACSJ/2016/25008

Editor(s):

(1) Anonymous.

Reviewers:

(1) Reddi Mohan Niadu Kalla, Pusan National University, Republic of Korea.

(2) Rachid Touzani, University Mohamed the First, Oujda, Morocco.

(3) Vishal W. Banewar, Government Vidarbha Institute of Science & Humanities, Amravati, India.

Complete Peer review History: <http://www.sciencedomain.org/review-history/16221>

Original Research Article

Received 12th February 2016
Accepted 9th May 2016
Published 16th September 2016

ABSTRACT

The synthesis of novel 1-amino-7-methyl-2,4,6,8,9-pent-azaphenoxazine (**13**) and 2-aldehyde-7-methyl-4-phenyl-3,4,6,8,9-pent-aza phenoxazine (**17**) is reported. The pent-aza compound (**13**) was prepared by suspending acetamide hydrochloride (**8**) in ethanol and treated with anhydrous hydrazine at 0°C to afford a pink solid compound acetamidrazone (**9**). Treatment of compound (**9**) with oxalyl chloride and heated at reflux, yielded 6-methyl-1,4,5-triazine-2,3-dione (**10**). This compound when reacted with 4,5-diamino-6-hydroxypyrimidine (**12**) in ethanol, provided a solid 1-amino-7-methyl-2,4,6,8,9-pent-azaphenoxazine (**13**) in excellent yield. The 2-aldehyde-7-methyl-4-phenyl-3,4,6,8,9-pent-aza phenoxazine (**17**) was obtained utilizing 3-aldehyde-5-amino-6-hydroxy-1-phenylpyridazine (**16**) which was produced by refluxing a mixture of furfural and phenylhydrazine in toluene. Treatment of compound (**16**) with 6-methyl-1,4,5-triazine-2,3-dione (**10**) in ethanol gave the pent-aza-phenoxazine (**17**) in good yield. These compounds were confirmed by IR, ¹H NMR, ¹³C NMR spectroscopy and MS spectrometry. The two novel products (**13**) and (**17**) were investigated for analgesic actions. The result compared favorably with three known analgesics: Paracetamol,

*Corresponding author: E-mail: odinem2005@yahoo.com;

Aspirin and Alabukun (a local analgesic). The doses of 0.125 to 0.150 g/kg injected into adult rabbits intraperitoneally lead to reduction of pain threshold. The result of the anti-inflammatory screening data revealed the dose and time dependant effect of these compounds in the carrageenan induced paw oedema. At time of 120mins, there was complete inhibition of oedema by 95.36% and 97.51% from the compounds **13** and **17** respectively, while the standard drug Zerodol (Aceclofenac) showed inhibition by 57.79%. The ability of these two compounds to antagonize carrageenan-induced oedema was correlated with anti-inflammatory potential.

Keywords: 1-amino-7-methyl-2,4,6,8,9-pent-azaphenoxazine; 2-aldehyde-7-methyl-4-phenyl-3,4,6,8,9-pent-azaphenoxazine; pain threshold; oedema; carrageenan-induced.

1. INTRODUCTION

Phenoxazine and its derivatives have not received a great deal of attention either theoretically or commercially. The only derivatives of commercial importance are a few dyes, known as the oxazine dyes [1,2,3]. Some of the oxazine dyes, particularly the Nile Blue dyes, are known to stain brain cancer tissue, while decrease the rate of growth of tumors [1]. The use of phenoxazine derivatives in biological staining is closely related to their use in the treatment of cancer and tuberculosis. Nile Blue A and Nile Blue 2R have been reported to retard growth of cancer [1,4,5].

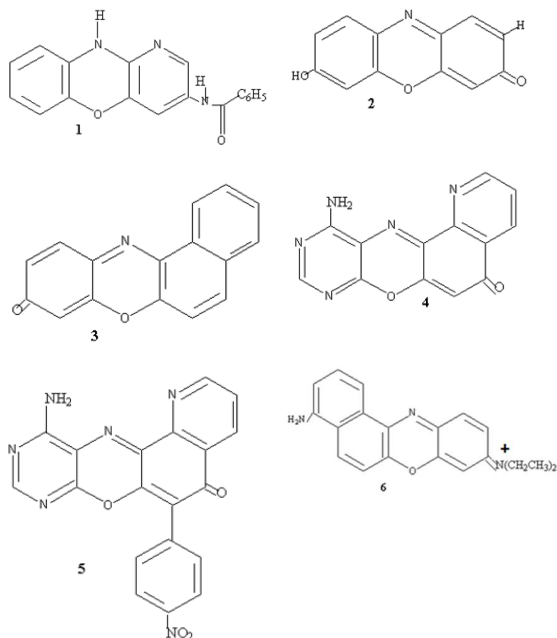
Phenoxazine derivatives have been reported for industrial application as acid-base indicator [6]. Phenoxazines containing pyrimidine nucleus have received significant pharmaceutical attention. Its ring is in clinical use such as anti-bacterial agents, anticancer, antiviral, antimalarial and antifungal agents [7].

Phenoxazine derivatives have been used also as antiperkinsonian, anticonvulsant, antihistaminic, antiviral and CNS depressants [7-13]. Phenoxazines have also been reported to be an effective multidrug resistant (MDR) modulator in cancer cells [7].

Phenoxazines containing pyridazine system, in recent times have received attention due to the biological activities. These pharmaceutical activities endowed in pyridazine have inspired chemists to synthesize phenoxazine systems containing pyridazine functionality in order to explore the usefulness of these heterocyclic template [14].

Phenoxazine systems of types (1): 3-benzamido azaphenoxazine [15]; (2): 7-hydroxy-phenoxazine-3one [16]; (3): Benzo[a]-phenoxazine-7-one[16]; (4): 11-amino-1,8,10-triazabenz[a]-phenoxazine-5-one [17]; (5): 6-(4-

nitrophenyl)-11-amino-1,8,10-triazabenz[a]phenoxazine-5-one [17]; (6): Nile Blue A [18] and their known pharmacological and industrial applications have previously been documented.



In continuation of our research programme directed towards the preparation of heterocycles with pharmaceutical recognitions [19,20], Pent-azaphenoxazines of types (13) and (17) to the best of our knowledge have not been reported.

In this paper we report the synthesis of these novel heterocyclic systems as a new group of anti-inflammatory heterocycles and their effect and other analgesics on pain.

2. MATERIALS AND METHODS

All chemicals used complied with international standards on health and safety as approved for commercial use by OECD (Organization of Economic Cooperation and Development),

UNEP (United Nation Environmental Programmes). This provides the environmental credentials of the chemicals. The melting points(m.p) were determined on a SMP3 melting point apparatus and were reported in °C uncorrected. Column chromatography was performed on Scharian Silica gel 60(70-230 mesh). Elemental analysis was performed using a Perking-Elmer 2400 CHN analyzer. The Infrared (IR) spectra were recorded in Cm^{-1} on a Bulk Scientific 500 Spectrophotometer. The ^1H NMR and ^{13}C NMR spectra were recorded on a Varian Gemini 2000 spectrophotometer operating 200 and 50 MH_z respectively. Chemical shifts were recorded as δ values in PPM referenced to the solvent. HPLC separations were performed on a Bulk scientific 500 apparatus using a reverse phase Lichrospher 100 RP-18(5M) column at room temperature (eluent: methanol/water 8:2,v/v).

2.1 Animals

In this study we followed the principle of laboratory animal care (NIH publication No 85-23, revised 1985). We employed adult rabbits of either sex weighing between 0.7-1.2 kg for the toxicity and analgesic studies, while the anti-inflammatory studies employed adult Wister rats 200-350g. All the animals were obtained from the local market in Gwagwalada, Abuja, Nigeria and maintained at the Animal Facility Centre of Kogi State University at standard conditions and temperature (25°C) and fed with standard diet (Pfizer feed, PLC, Lagos) and *tridaxprocuben*, carbage and water for one week.

2.2 Synthesis

The synthetic routes for the two novel compounds are outlined in schemes 1 and 2. The details are given below:

2.3 Acetamidine Hydrochloride (8)

This compound was prepared as in the literature [21]. Solid acetamido ethyl ether hydrochloride (98 g, 3 mol) was broken up and transferred to a dry mortar. 20 ml of super dry ethanol was added to it and grounded to paste before it was returned to the reaction vessel. Alcoholic ammonia solution (700 ml) was added to the contents in the vessel and Stirred mechanically for 4h, filtered and the filtrate was 2389(CH_3). ^1H NMR: 5.71 (m, NH_2 protons), 5.76 (m, CH_3). ^{13}C NMR:

164 (C-NH), 139 (C=N), 89.5 (CH_3), 88.9 (C-H). Calculated for $\text{C}_2\text{H}_8\text{N}_2\text{Cl}$: C, 25.25; H, 8.48; N, 29.45:C.

2.4 Acetamidrazone (9)

9.5g, 1 mol of acetamidine hydrochloride (8) was suspended in super dry ethanol (50 ml) and treated with 3.2 g anhydrous hydrazine at 0°C for four minutes. The ethanol content was evaporated at 35°C to give a pink solid. M.p:261-265°C. Yield:30g, 91%. IR: 1446 (C=N), 817 (C-N), 3284-3180 (N-H), 2388 (CH_3 group), ^1H NMR: 5.70 (m. NH_2 protons), 5.75 (m. CH_3). ^{13}C NMR: 163 (C-NH), 138 (C=N), 89.4 (CH_3), 88.8 (C-H). Calculated for $\text{C}_2\text{H}_7\text{N}_2$: C, 40.64; H, 11.95; N, 47.41%. Found: C, 40.63; H, 11.93; N, 47.40%.

2.5 6-methyl-1,4,5-triazine-2,3-dione (10)

The synthetic route followed a modification of the methods already described by Naidu et al [22,23]. 0.03 mol, 2.18 g of acetamidrazone (9) was suspended in super dry ethanol (40 ml) and treated with oxalyl chloride (0.03 mol, 3.96 g) and heated while refluxing for 2 h. The reaction mixture was allowed to cool and filtered. The resulting solid was crystallized from ethanol/water. M.p:203-206°C. Yield:22g, 79%. IR: 1556 (C=O), 1585 (C=N), 1170 (C-N), 3235 (N-H), 2389 (CH_3). ^1H NMR: 6.8 (s, 1H, NH), 7.6 (4H, s, NH), 5.73 (s, CH_3). ^{13}C NMR: 187.63 (C=O triazine); 182 (s, C6); 27.28 (CH_3); 171.30 (C-NH); 130.73 (C=N). Calculated for $\text{C}_4\text{H}_5\text{N}_3\text{O}_2$: C,37.78; H, 3.97; N, 33.06; O, 25.19% Found: C, 37.76; H, 3.46; N, 33.04; O, 25.17%.

2.6 1-amino-7-methyl-2,4,6,8,9-pent-azaphenoxazine (13)

0.1 mol, 12.7 g of 6-methyl- 1,4,5-triazine-2,3-dione (10) was mixed with 0.1 mol, 21.7 g 4,5-diamino-6-hydroxypyrimidine (12) in ethanol (100 ml) and heated at reflux for 3 h. The reaction mixture was cooled and the resulting solid filtered. The product was crystallized from ethanol/water. M.p:189°C. Yield: 32 g, 98%. IR: 3370 (N-H phenoxazine), 690 (C-O-C), 1439 (C=N0), 817 (C-N). ^1H NMR: 7.9 (1H, s,N-H phenoxazie), 5.79 (CH_3), 5.73 (s, NH_2 protons). ^{13}C NMR: 130.1 (C=N, C4), 131.20 (C=N, C8), 28.11(m, CH_3), 138.1(C- NH_2). Calculated for: $\text{C}_8\text{H}_7\text{N}_7\text{O}$: C, 14.19; H, 3.25; N,45.13; O,7.37%. Found: C,14.19; H, 3.24; N, 45.13; O, 7.36%.

2.7 3-aldehyde-5-amino-6-hydroxy-1-phenylpyridazine (16)

This was prepared using a modification of the methods already described by Dilek et al. [14] and Alberto et al. [24]. A mixture of 4-amino-5-hydroxyfurfural (1.7 ml 0.02 mol) (**14**) and phenylhydrazine 92.89 g, 0.02 mol) in toluene (20 ml) was heated for 4 h. The solvent (toluene) was removed by evaporation and the oily residue was treated with diethyl ether overnight. The crude product formed was crystallized from acetic acid. M.p: 356-358°C. Yield: 58%. IR: 3341 (N-H pyridazine); 816 (C-N); 1722, 1654 (C=O); 1382 (C=C arom.); 2408 (C-H arom.); 3430 (OH); 1631 (ArH); 3291 (NH₂); 658 (C-O). ¹H NMR: 5.39 (s, NH₂); 6.70 (OH); 7.54-7.97 (Ar-H); 5.29 (s, NH pyridazine); 6.7 (C-H). ¹³C NMR: 138.3 (C-NH₂); 175.6 (s, C4); 137.28-130.20 (m, Ar.C); 129.6 (C-N, C6); 138.3 (C-NH₂); 192.28 (C-N, C6); 192.28 (C=O pyridazine); 78.9 (CH); 112.40 (C=C). Calculated for C₁₁H₁₁N₃O₂: C,60.80; H, 5.11; N, 19.35; O, 14.74%. Found: C,60.79; H,5.10; N,19.35; O,14.73%.

2.8 2-aldehyde-7-methyl-4-phenyl-3,4,6,8,9-pent-azaphenoxazine (17)

0.1 mol, 12.7 g of 6-methyl-1,4,5-triazine-2,3-dione (**10**) was mixed with 0.1 mol, 32 g 3-aldehyde-5-amino-6-hydroxy-1-phenylhydrazine (**16**) in ethanol (100 ml) and heated at reflux for 3 h. The reaction mixture was cooled, the solid formed was filtered and crystallized from ethanol/water. M.p: 192°C, Yield: 41g.76%. IR: 3451 (N-H pyridazine); 3427 (N-H phenoxazine); 1631 (Ar-H); 1444 (C=N); 816 (C-N); 657 (C-O); 1651 (C=O); 2001 (C-H arom.); 1376 (C=C arom.); 658 (C-O phenoxazine); 2385 (CH₃ groups). ¹H NMR: 8.1 (s, NH phenoxazine); 6.9 (m, ArH); 5.36 (N-H pyridazine); 2.97 (s, CH₃). ¹³C NMR: 186.63 (s, C1); 128.30-134.17 (m, Ar.C); 130.6 (C=N); 131.6 (C=C); 133.8 (C=O). Calculated for: C₁₅H₁₂N₆O₂: C, 58.42; H, 3.93; N, 27.26; O,10.39%. Found: C, 58.41; H, 3.92; N, 27.24; O, 10.37%.

2.9 Determination of LD₅₀

The acute toxicity (LD₅₀) of the novel phenoxazines (**13 & 17**) was determined in adult rabbits using the methods of Lorke [25], Amos et al. [26], Azuine et al. [27], Gurad et al. [28], and Odin et al. [20]. The animals were divided randomly into eight groups of five rabbits each. Widely differing doses of 10, 100, 1000, 1500, 2000, 3500 and 5000 mg/kg of the compounds

were administered intraperitoneally and orally on groups 1-7. The eighth group was administered normal saline (10 ml/kg) and served as control. The animals were monitored for 72 h for signs of toxicity such as spontaneous motor activity, exploratory activity, climbing behavior, paw-licking, convulsions, lacrimation and salivation [19,29-31]. At the end of the experiment, they were sacrificed and then autopsied and examined microscopically for any pathological changes. Pieces of organs, such as liver, heart and kidney were examined for histopathological changes.

2.10 Haematological Parameters

The effect of the synthesized compounds (**13 & 17**) on haematology of adult rabbits after the administration of the compounds on day 3, (72nd h) was determined. This includes haematological parameters such as Packed cell volume (PCV), haemoglobin (Hb), red blood cells (RBC) and white blood cells (WBC). The methods of McDonard et al. [32]; Subhasini et al. [33]; Olamide and Mathew, [29] were employed and modified. The RBC, WBC, Hb and PCV were measured by an auto Hematology Analyzer. These parameters were quantified in each pent-azaphenoxazine compound (**13 & 17**) and control. The results are presented in Table 2.

2.11 Test for Analgesia

The hot plate method of Onabanjo et al. [34] was modified thus: The temperature of the flask placed in water-bath was maintained in the range of 55-60°C. The rabbits were shared into twelve groups of four each. Groups **1 & 2** were administered intraperitoneally 3 ml and 5 ml normal saline respectively, and served as controls. Groups **3 & 4** were administered 0.125 and 0.150 g/kg doses of aspirin respectively. Groups **5 & 6** had 0.125 and 0.150 g/kg doses of paracetamol respectively, while groups **7 & 8** were given 0.125 and 0.150 g/kg doses of alabukun respectively. The chosen concentrations were based on toxicity studies. Animals in groups **9 & 10** were injected with 0.125 and 0.150 g/kg compound **13**, while those in groups **11 & 12** were also injected with 0.125 and 0.150 g/kg compound **17**. All the treated animals were left for 30 min for drug digestion. They were then individually placed into the flask and the reaction time was monitored. The reaction time was recorded as the jump latency [35]. Time of maximum intolerance to the heat was regarded as the maximum analgesic period (Table 1).

2.12 Anti-inflammatory Studies

We employed the modified methods of Winter et al. [36], Azuine et al. [27], Ratheesh and Helen, [37], Adeyemi et al. [38] and Haruna and Cyril, [39]. Wister rat comprising male and female weighing between 180-300 g were divided into six groups of five rats each.

Inflammation of the right hind paw was induced by injecting 1% carrageenan suspension in 0.9% NaCl solution into the sub planter surface. Animals in group 1 received normal saline (20 ml/kg) and were labeled as negative control. Group 2 and 3 animals were administered intraperitoneally compound 13 doses of 50 mg/kg and 100 mg/kg respectively. The fourth and fifth group animals were administered compound 17 doses of 50 mg/kg and 100 mg/kg respectively while group six animals were given zerodol (aceclofenac-50 mg/kg) injected intraperitoneally and were designated as positive control. All the drugs were administered 60 minutes before the subplanter injection of the phlogistic agent. The paw volume was measured after every 20 minutes for a period of 120 minutes by a volume displacement methods [27] using a digital plethysmometer, Coulbourn instrument-AHarvard apparatus company (Table 3).

The average inflammation, percentage inflammation and percentage inhibition of

Oedema were calculated on each dose and recorded (Table 4, 5, and 6). The average inflammation was calculated according to the methods of Azuine et al. [27]: $L_t - L_0$ where L_t is the linear circumference at the time t and L_0 is the linear circumference at zero time (Table 4).

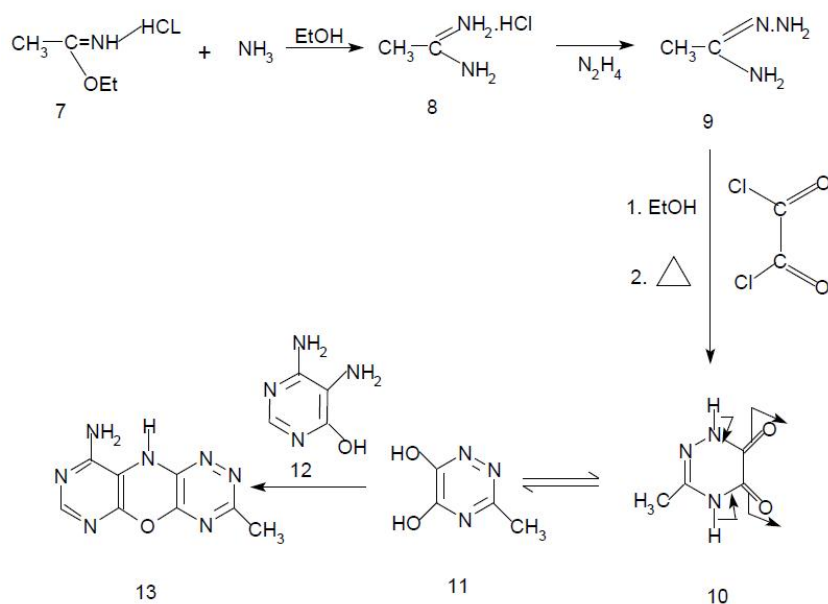
The percentage inflammation was also calculated according to the method of Azuine et al. [27]:

$A/B \times 100$ where A is the average inflammation of treated group at time t while B is the average inflammation of the control at the same time (Table 5). The percentage inhibition of Oedema (Table 6) is calculated as follows:

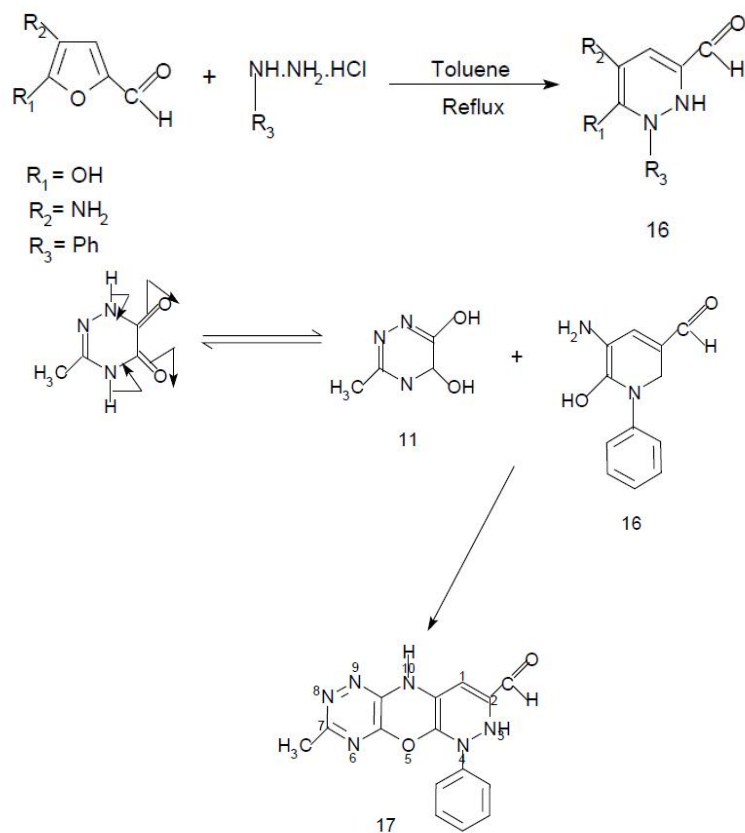
$100 - \text{percentage inflammation.}$

3. RESULTS

Acetamidine hydrochloride (8) was prepared from solid acetamido ethyl ether hydrochloride in super dry ethanol followed by the addition of alcoholic ammonia. Treatment of this compound (8) with anhydrous hydrazine afforded a pink solid acetamidrazone (9). 6-methyl-1,4,5-triazine-2,3-dione(10) was produced by suspending compound (9) in super dry ethanol treated with oxalyl chloride and refluxed for 2h.



Scheme 1



Scheme 2

Compound (10) when mixed with 4,5-diamino-6-hydroxypyrimidine in ethanol yielded a solid 1-amino-7-methylpent-azaphenoxazine(13) (scheme 1). Compound (16), 3-aldehyde-5-amino-6-hydroxyl-1-phenylpyridazine was synthesized by heating a mixture of 4-amino-5-hydroxyfurfural and phenylhydrazine in toluene. The crude product was crystallized from acetic acid. When this compound (16) was mixed with 6-methyl-1,4,5-triazine-2,3-dione(10) in ethanol and crystallized in ethanol/water, the result was a solid compound 2-aldehyde-7-methyl-4-phenylpent-azaphenoxazine(17) (scheme 2).

The structural assignment of the synthesized compound is based on the spectral data. In the IR spectrum of compound (8), the hydrogen bonded N-H stretching appeared at 3283-3179 cm^{-1} . Absorption band at 2389 cm^{-1} represent CH_3 groups. The C=N absorption band appeared at 1445 cm^{-1} , while that at 816 cm^{-1} is characteristic of C-N stretching. 1H and ^{13}C NMR. Studies of this compound(8) confirmed the structure. In 1H NMR spectra data, compound (8) showed multiplet at δ 5.76. In ^{13}C NMR spectrum,

a characteristic signal appeared for C-N in the range of δ 164 while the signal for C=N, (CH_3) and (CH) appeared in the range of δ 139, δ 89.5 and δ 88.9 respectively.

The IR spectrum of compound (9) showed broad band at 3284-3180 cm^{-1} for hydrogen bonded N-H stretching. The bands 1446 cm^{-1} and 817 cm^{-1} were noticed for (C=N) and (C-N) stretching respectively, while that for CH_3 stretching appeared at 2388 cm^{-1} . In the 1H NMR spectral data, compound(9) showed multiplet at δ 5.70 and δ 5.75 due to NH_2 and CH_3 protons. A characteristics signals appeared for (C-NH), (C=N) (CH_3) and (C-H) in the range of δ 163, δ 138, δ 89.4 and δ 88.8 respectively in the ^{13}C NMR spectrum.

In the IR spectrum of compound (10), a broad band for hydrogen bonded NH stretching showed at 3235 cm^{-1} . The band at 2556 cm^{-1} indicated C=O stretching while the band at 1585 cm^{-1} and 1170 cm^{-1} were for (C=N) and (C-N) respectively. The absorption at 2389 cm^{-1} were located for CH_3 groups.

In the ^1H NMR spectrum of this compound, the singlet for 2NH proton appeared in the region δ 5.80 and δ 7.6, while the CH_3 protons showed a singlet in the region δ 5.73. The ^{13}C NMR studies of this compound confirmed the structure. A characteristic signal appeared for (C=O triazine) at δ 183.67, while that of C=N were seen in the range δ 130.73. The signal at δ 27.78, δ 171.30 and δ 130.73 were located for (CH_3), (C-NH) and (C=N) respectively.

In compound (**13**) the band at 3370 cm^{-1} is assigned to N-H phenoxazine while the absorption band at 690 cm^{-1} , 1439 cm^{-1} and 817 cm^{-1} are characteristic of C-O-C, C=N and C-N stretching vibration. In the ^1H NMR spectrum, the singlet for NH, CH_3 and NH_2 protons appeared at δ 7.9, δ 5.79 and δ 5.73 respectively. In the ^{13}C NMR spectrum, a characteristic signals appeared for (C-NH, C_4) and (C=N, C_8) in the range of δ 130.1 and δ 131.20 respectively, while that of (CH_3) and (C-NH $_2$) were detected in the range of δ 28.11 and δ 138.1 respectively.

In the IR spectrum of compound (**16**), the absorption band at 3341 cm^{-1} and 3291 cm^{-1} represent the hydrogen bonded N-H pyridazine and NH_2 stretching vibration. There were a number of peaks at 816 cm^{-1} , 1722 cm^{-1} , 1654 cm^{-1} , 1382 cm^{-1} , 2408 cm^{-1} , 3430 cm^{-1} and 658 cm^{-1} representing C-N, C=O, C=C, OH and C-O respectively. The band at 1631 cm^{-1} is assigned to Ar-H stretching. In the ^1H NMR spectral data, compound(**16**) showed a singlet at δ 5.39 and δ 5.29 due to NH_2 and N-H protons. The Ar-H protons appeared in the region of δ 7.54 while the OH protons in the compound were located in the range of δ 6.70. In ^{13}C NMR spectrum, a characteristic signal appeared for (C-NH $_2$), (C_4), (C-N, 6), (C-NH $_2$), (C=O pyridazine), (CH) and (C=C) in the range of δ 138.3, δ 175.6, δ 129.6, δ 138.3, δ 192.28, δ 78.9, and δ 112.40 respectively, while the (Ar-C) were in the range of δ 137.28- δ 3130.20.

The IR spectrum of compound (**17**) showed absorption bands at 4351 cm^{-1} and 3427 cm^{-1} representing the hydrogen bonded N-H pyridazine and N-H phenoxazine respectively. The absorption band at 1631 cm^{-1} was for Ar-H stretching, while 1444 cm^{-1} and 816 cm^{-1} represent (C=N) and (C-N) stretching respectively. There were other peaks at 657 cm^{-1} , 1651 cm^{-1} , 2001 cm^{-1} , 1376 cm^{-1} , 658 cm^{-1} , and 2385 cm^{-1} which were for C-O, C=O, stretching, aromatic C-H, aromatic C=C, C-O phenoxazine and CH_3 groups. In the ^1H NMR spectrum,

compound (**17**) showed singlet for N-H phenoxazine, $-\text{CH}_3$ protons at δ 8.1 and δ 2.97 respectively and multiplet for Ar-H at δ 6.9, while the N-H pyridazine protons were located at δ 5.36. In the ^{13}C NMR spectrum, some signals were noticed at δ 186.63, δ 130.40, δ 131.6 and δ 133.8 for (s, C I), C=N, C=C, and C=O respectively. That of Ar.C were detected at δ 128.30 -134.17.

The mass spectrophotometric studies performed on the synthesized compounds confirmed the molecular weight values. The result of the anti-inflammatory test on the novel heterocyclic systems and their effect and other analgesics on pain are as presented in Tables 1,2,3,4,5 and 6.

4. DISCUSSION

From the result of the acute toxicity (LD_{50}) of the novel phenoxazine compounds (**13** & **17**), differing doses of 10 mg/kg to 5000mg/kg of the compound were selected so that the entire range of toxicity from high acute toxicity to virtual non-toxicity could be tested. The weight of the animals after the test showed that they all gained weight. This was taken as a sign of having survived the acute intoxication. The mice treated intraperitoneally up to 2000 mg/kg did not show sign of toxicity compared with control animals. Similarly no significant effects was detected in animal treated orally with the compounds up to 5000 mg/kg. This high dose only produced intense quietness. This effect was reversed within 3 hrs. All the animals survived the test. That is no death was recovered. This clearly demonstrate that the two phenoxazine compounds (**13** and **17**) even at high dose of 5000 mg/kg (5 g/kg) were non-toxic.

The haematological profile of the adult rabbits were affected by the two phenoxazine compounds. The WBC protect the body against foreign invaders such as bacteria. Therefore a low WBC count usually indicates that the neutrophile count is low and it becomes easier to get an infection and harder to recover from it. A low number of WBC is called Leukopenia. A WBC count below 4500 is below normal. The normal number of WBC in the blood is 4,500-10,000 per microlitre (mcl). From Table 2, it is observed that the number of WBC in the blood of the treated rabbits compared favourably with control treated animals. The observed significant increase in values of WBC at 1500-5000 mg/kg may be attributed to stimulation of the immune system. This significant in dose dependent manner can be an indication of the non-toxicity in

high doses. The compound therefore might be useful in the management of the inflammatory and tumor related disease.

The body needs oxygen to release energy and keep organs and tissues healthy. This is one of the functions of the RBC using a substance called haemoglobin (hb). A person with a low Hb level may have anemia [40]. A normal Hb level is between 12% and 16% while a normal RBC count is between 4.7-9.4 microlitre (mcl).

From Table 2, the significant decrease in the level of haemoglobin concentration and increase in the level of red blood cell count with increasing doses are within the normal level. This is an indication that the novel phenoxazine compounds cannot cause anemia in animals.

The PVC values is a measure of the oxygen carrying capacity of the blood. Table 2 revealed a decrease in the PVC values with increasing doses, indicating the degree of stress on the test animal health [41].

Results in Table 1 indicate that the pent-azaphenoxazines (**13** and **17**) administered to the rabbit were highly active compared to the human analgesic drugs employed in the study considering the mean reaction times. At a considerable concentration of 0.150 g/kg, the decreasing order of action of the human analgesics were alabukun, paracetamol and aspirin. At a lower concentration of 0.125 g/kg, compounds **13** and **17** compared favourably with the three human analgesics. This implies therefore that the active principle are equally present in the synthesized compounds. The doses of 0.125 to 0.150 g/kg injected into the adult rabbits intraperitoneally resulted in reduction of pain threshold. This level of activity is remarkable. It was previously observed that

aspirin has a very poor analgesic action through the inhibition of prostaglandin synthesis [42]. This observation is supported by the result obtained in the study at aspirin concentration of 0.150 g/kg and the result of aspirin versus saline applications (Table 1).

The effect of the pent-azaphenoxazine compounds (**13** and **17**) on carrageenan-induced oedema in rats are shown in Tables 3,4,5 and 6.

From Table 4 it can be seen that in control animals, the sub-planter injection of carrageenan produced local oedema after 20 min. The two compounds at 50 mg/kg and 100 mg/kg demonstrated a significant anti-inflammatory effect. The dose dependent effect of these compounds in the carrageenan induced paw oedema showed that the effect was real and not due to counter irritant activity. Table 4 also revealed that apart from being dose and time dependent, the action of pent-azaphenoxazine compounds on carrageenan induced oedema also depends on the nature of the di-azine molecules and the type of substituent available on them (compound **13** and **17**). The average inflammation was 0.29 mm and 0.09mm when the rat was pretreated with 50 mg/kg and 100 mg/kg respectively with compound (**13**) in the first 20 min, while that of compound **17** was 0.28 mm and 0.08 mm.

From Table 6, at time 120 min there seems to be a complete inhibition of oedema by 95.36% and 97.51% for compounds **13** and **17** respectively. The standard drug Zerodol (Aceclofenac) at 50 mg/kg showed inhibition by 57.79% (Table 6).

This significant changes was probably due to the nature of the di-azine molecule in the pent-azaphenoxazine systems and the type of substituent available in them.

Table 1. Analgesic property of the pent-azaphenoxazines

Group	Test agent	Dose(g/kg)	Mean reaction time (sec)
1	Saline	-	12±6.32
2	Saline	-	13±8.23
3	Aspirin	0.125	14±6.6
4	Aspirin	0.150	16±2.32
5	Paracetamol	0.125	37±4.32
6	Paracetamol	0.150	40±3.12
7	Alabukun	0.125	42±4.52
8	Alabukun	0.150	48±4.52
9	Pent-azaphenoxazine(13)	0.125	69±14
10	Pent-azaphenoxazine(13)	0.150	73±20
11	Pent-azaphenoxazine(17)	0.125	72±11
12	Pent-azaphenoxazine(17)	0.150	78±64

Table 2. Effects of pent-azaphenoxazine (13 and 17) on haematology of adult rabbits

Parameters	Compound 13 group							Control	Compound 17 group						
	1	2	3	4	5	6	7		1	2	3	4	5	6	7
RBC(mm ³)	8.35±0.03	8.40± 0.36	8.46±0.94	8.59±0.49	8.68±0.93	8.89±0.36	9.12±0.37	9.28± 0.11	8.38±0.17	8.36±0.11	8.43±0.11	8.56±0.32	8.66±0.18	8.87±0.61	9.01±0.46
WBC(mm ³)	6375±0.11	6492±0.30	7742±35	7901±72	8.058±0.21	8.079±0.86	8148±0.61	6580±41	6381±0.91	6499±0.12	7811±10	7981±33	8073±0.20	8106±	8192±
Hb(%)	14.3±0.37	13.9±0.14	13.3±0.58	12.7±0.34	12.4±0.71	12.09±0.80	11.7±0.43	15.9±0.19	14.01±0.42	13.6±0.18	13.09±0.64	12.5±0.16	12.2±0.93	12.0±0.11	11.5±0.36
PCV(%)	52.3±19	50.5±0.13	47.2±22	44.3±0.19	40.7±89	39.6±75	39.42±77	58.4±0.14	51.90±11	49.3±0.71	46.9±0.38	43.1±0.34	39.9±0.31	38.0±0.16	38.6±0.14

Table 3. Paw volume (mm³)

Treatment group	0 min	20 min	40 min	60 min	80 min	100 min	120 min
Normal saline (20 ml/kg)	0.50	1.08±0.07	1.13±0.10	1.14±0.03	1.05±0.02	1.03±0.04	1.01±0.12
Zerodol (aceclofenac) 50 mg/kg	0.53	0.85±0.03	0.86±0.13	0.84±0.17	0.78±0.72	0.77±0.10	0.75±0.19
Compound 13 (50 mg/kg)	0.55	0.84±0.41	0.85±0.11	0.84±0.01	0.78±0.14	0.77±0.00	0.78±0.04
Compound 13 (100 mg/kg)	0.63	0.72±0.06	0.71±0.31	0.70±0.47	0.68±0.13	0.67±0.11	0.65±0.10
Compound 17 (50 mg/kg)	0.56	0.84±0.20	0.84±0.17	0.82±0.16	0.78±0.48	0.76±0.13	0.74±0.91
Compound 17 (100 mg/kg)	0.69	0.72±0.13	0.71±0.11	0.69±0.51	0.68±0.00	0.66±0.14	0.65±0.00

Table 4. Average inflammation (mm) of the right hind paw

Treatment group	20 min	40 min	60 min	80 min	100 min	120 min
Normal saline (20 ml/kg)	0.581	0.63	0.64	0.55	0.53	0.54
Zerodol (aceclofenac) 50 mg/kg	0.32	0.33	0.31	0.25	0.24	0.22
Compound 13 (50 mg/kg)	0.29	0.30	0.29	0.23	0.22	0.20
Compound 13 (100 mg/kg)	0.09	0.08	0.07	0.05	0.04	0.02
Compound 17 (50 mg/kg)	0.28	0.28	0.26	0.22	0.20	0.18
Compound 17 (100 mg/kg)	0.08	0.07	0.05	0.04	0.02	0.01

Table 5. Percentage inflammation (%) of right hind paw

Treatment group	20 min	40 min	60 min	80 min	100 min	120 min
Zerodol (aceclofenac) 50mg/kg	54.89	51.88	48.50	46.18	46.00	42.21
Compound 13 (50mg/kg)	49.87	47.81	44.75	42.09	41.40	39.83
Compound 13 (100mg/kg)	15.40	12.58	11.43	9.30	7.19	4.64
Compound 17 (50mg/kg)	48.32	43.88	44.32	39.16	38.44	36.15
Compound 17 (100mg/kg)	14.25	11.84	8.47	6.82	4.71	2.40

Table 6. Percentage inhibition of oedema (%)

Treatment group	20 min	40 min	60 min	80 min	100 min	120 min
Zerodol (aceclofenac) 50 mg/kg	45.11	48.12	51.5	53.82	54.00	57.79
Compound 13 (50 mg/kg)	50.13	52.49	55.25	57.91	58.60	60.17
Compound 13 (100 mg/kg)	84.60	87.42	88.57	90.70	92.70	95.36
Compound 17 (50 mg/kg)	51.68	56.12	58.68	60.84	61.56	63.85
Compound 17 (100 mg/kg)	85.75	88.16	91.56	93.48	95.29	97.51

Compound **13** contains a pyrimidine structure with an amino substituent. Phenoxazines containing pyrimidine nucleus have received significant pharmaceutical attention [7]. This ring has aided the clinical use of phenoxazines as anti-cancer, anti-malarial, antibacterial agents, anti-viral and anti-fungal agents [7].

Compound **17** contains Pyridazine nucleus, one activating group and one carbonyl group that deactivates the pyridazine ring. Phenoxazines containing pyridazine systems, in recent times have been reported to possess biological activities [14]. These properties including the pyridazine functionality present in compound **17** probably have enhanced the percentage inhibition of this compound above that of compound **13**.

The result of this work as indicated in Tables 1,2,3,4,5 & 6 clearly demonstrate the significant anti-inflammatory properties of these novel pent-azaphenoxazine compounds (**13 & 17**), their effect and other analgesics on pain.

The suppression of oedema by the compounds may be due to the presence of the diazine systems.

4.1 The Mode of Action

The carrageenan induced oedema in rats is due to the release of histamine, serotonin and prostaglandin [37,43]. The two pent-

azaphenoxazines (**13 & 17**) including the control drug Zerodol (Aceclofenac) are among the NSAIDs (Non steroidal anti-inflammatory drugs). The mechanisms of action of these systems may be due to their ability to irreversibly inhibit prostaglandin G/H synthesis by acting on the active site of the enzyme. They prevent the formation of prostaglandin synthase including thromboxane, prostacyclin and other prostaglandins [44]. When a tissue is injured or stimulated, prostaglandin synthesis in that tissue increases. The prostaglandins are mediators of inflammation and they also sensitize nerve endings, lowering their threshold of response to stimuli and the tenderness of inflammation [44].

The fact is that a drug that prevents the synthesis of prostaglandins is likely to be effective in relieving pains due to inflammation of any kind. This is probably how Zerodol (aceclofenac) and other non steroidal anti-inflammatory drugs (NSAIDs) act [43]. Meaning that NSAIDs act by inhibiting cyclo-oxygenase (prostaglandin G/H synthase). This shows that the synthesized compounds (**13 & 17**) will relieve pain when there is some tissue injury with consequent inflammation.

5. CONCLUSION

Azaphenoxazines as anti-inflammatory drugs has long been recognized. Compounds 13 and 17 are the first pent-azaphenoxazines to possess

anti-inflammatory properties. The anti-inflammatory screening experiments of the two compounds on the carrageenan induced paw oedema is a measure of the anti-inflammatory activity. Complete inhibition of oedema by 95.36% and 97.51% from these compounds revealed their anti-inflammatory effects. This procedure has been accepted as a parameter for evaluating inflammatory conditions. The ability of these compounds to antagonize carrageenan-induced oedema in Wister rats has been correlated with anti-inflammatory potential.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

- Leonard OM. Reactions of phenoxazine and some of its derivatives. A dissertation submitted to the graduate faculty in partial fulfillment of the requirement for the degree of Doctor of Philosophy. Iowa State College; 1957.
- Venkataraman VK. The chemistry of synthetic dyes, New York, Academic press Inc. 1952;1:xvi.
- Lubs HA. The chemistry of synthetic dyes and pigment. Reinhold Publishing Cooperation, N.Y; 1955.
- Lewis RM, Slovitor HA, Goland PP. In vivo staining and retardation of growth of sarcomata in mice. Anal. Record. 1946;95:89.
- Riley JF. Retardation of growth of a transplantable carcinoma in mice fed basic metachromic dyes. Cancer Research. 1948;8:183.
- Toba OT, Chris OU, Izuchukwu UD. Synthesis of 1-azabenz[a]phenoxazine-5-one and 11-amino-1,5,10-triazabenz[a]phenoxazine-5-one and their functionalized aryl derivatives via mizoroki-Heck arylation methodology. Orient J. Chem. 2014;31(1): 371-378.
- Idries M, Marshkov A. Synthesis of new [10H-substituted phenoxazine-3-YL]-6-pyrimidine-2-phenyl (thiol/oL/amine/thiol) pyrocoles. Thi-Qur Med J. 2010;4:120-126
- Harwood PD, Jestad AC. J. Parasitol. 1938;24:16-18.
- Halpem BN. J. Am. Med. Assoc. 1990; 129:1219-1222
- Kalinowasky LB, Hoch PH. Somatic treatment in psychiatry. Academic Press New York. 1961;122-132.
- Clane NT, Witten LK, Eilmer DS. Aust, Ver. J. 2003;23:344-346.
- Douglas JR, Baker NF. J. Am. Vet. 1992; 17:318-320.
- Janssen PAJ, Niemegeers CJE. Arzneim Forsch. 1996;15:1196-1199.
- Dilek U, Emin S, Yunus A. A new method for the preparation of pyridazine system; Experimental data and semi-empirical PM3 calculation. Turk. J. chem. 2006;30:691-701.
- Agbo SA, Igbum GO, Anoh VA, Swande PI. Synthesis and *In Vitro* antimicrobial activity of some novel azaphenoxazine carboxamide derivatives. 10SR J. Applied Chem. 2015;8(4):21-25.
- Olga W, Joseph M, Gurnner W, Kristin S, Masami Imre O, Nobru M, Kristyna M. Benzo [a] phenoxazine. A new group of potent p-glycoprotein inhibitors. *In vivo*. 2006;20:109-114.
- Quambe LQ, Pare B, Jonnalagadda SB. kinetics and mechanism of reaction of acidic chlorites with phenoxazine dyes, Nice blue and melalola's blue. Bull. chem. soc. Ethopia. 2005;19(1):103-116.
- Odin EM, Onoja PK, Sale JF. Synthesis, characterization and neuropharmacological activity of novel angular pentacyclic phenothiazine. Int. J. Phy. Sci. 2013; 8(26):1374-1381.
- Odin EM, Onoja PK. Synthesis of bridge head-fused 1,2,3-Triazol[1,5-c]-1,2,4-Triazines. Novel Anti-inflammatory and analgesic therapeutic systems. Global J. Sc. frontier research in chemistry. 2015; 15(3)1:19-33.
- Dox AW, Whitmore FC. Acetemidine Hydrochloride. Org. Synthesis. 1941;1:5, 1928;8:1.
- Naidu KRM, Kalivulla SA, Rassheed S, Fakurazi S, Aruselvan P, Lasekan O, Abas F. Synthesis of bisindolyl methanes and their Cytotoxicity Properties. Int. J. Mol. Sci. 2013;14:1843-1853.
- Naidu KRM, Krishna BS, Kumar MA, Aruselvan P, Kalivulla SA, Lasekan O. Design, synthesis and antiviral potential of 14-Aryl/Heteroaryl-14H-dibenzo[a,j] Xanthenes using an efficient Polymer supported Catalyst. Molecules. 2012;19: 7543.
- Alberto C, Eddy S, Enrique R. Pyridazine derivatives. Part 33: Sonogashira approach in the synthesis of 5- substituted -6-phenyl-3(2H)-pyridoquines. Tetrahedron 2003;59:2477-2484.

24. Lork D. A new approach to practical acute toxicity. *Achieves of Toxicity*.1923;54:257.
25. Amos S, Binda H, Vongtan B, Chindo J, Abba N, Odin EM, Okwute SK, Akah P, Wambebe C, Gamaniel K. Sedative effect of the methanolic leaf extract of *Newbouldia laevis* in mice and rats. *Boll Chin Farmac*. 2002;144(6):471-475.
26. Azuine MA, Ibrahim K, Ewerem NM, Wambebe C. Anti-inflammatory and anti-cancer activity of *Newbouldia laevis* extract in rats and mice. *J. Pharm. Research and Development* .1996;1(16).
27. Gurad A, Anshoo G, Pravin K, Abadesh K. Acute toxicity studies. A safer and more effective analogous of N,N-diethylphenyl acetamide. *J. Med. Enthanol*. 2011;48(6): 1160-1166.
28. Olamide EA, Mathew OA. Phytochemical and acute toxicity of ethanolic extract of *Enatia chlorentha (oliv)* stem bark in albino rats. *Int. Toxicol*. 2013;6(3):145-151.
29. Salawu OA, Chindo BA, Tijani AY, Obiadike IC, Salawu TA, Akingbosote AJ. Acute and sub-acute toxicological evaluation of the methanolic stem bark extract of *Crossopteryx febrifuga* in rats. *Afri. J. Pharm. Pharmacol*. 2009;3(12):621-626.
30. Karam JA, Suatek RJ, Karakiewicz PI. Use of preparative plasma endolin for prediction of lymph node metastasis in patients with clinically localized prostate cancer. *Clin. Cancer Res*. 2008;14(5): 1418-1422.
31. McDonald GA, Dodds TC, Cruickshanks B. *Atlas of Haematology*. Churchill Living Stone. Edinburgh London and New York. 1978;309.
32. Subhasini S, Kalpana A. Schweter V, Sharma KP. Toxicity assessment of textile dyes waste water using swiss albino rats. *Austri. J. Ecotex*. 2007;81-85.
33. Onabanjo AO, John TA, Sokande AA, Samuel OT. Analgesic and anti-inflammatory effects of *Chesmanthera dependens*. *Int. phern. Biology*. 1991; 29(1):24-28.
34. Agbaje EO, Onabanjo AO. Analgesic and antipyretic action of *Enatia chlerantia* extract in some laboratory animals. *Nig. J. Nat. Prod. Med*. 1998;2:24-25.
35. Winter A, Risley EA, Nuss GW. Carrageenan induced Oedema in the hind paw of rat as an assay for anti-inflammatory activities. *Proc. soc. Exp. Biol. Ther*. 1962;111:544-547.
36. Ratheesh M, Helen A. Anti-inflammatory activity of *Ruta graveolens linn* on carrageenan induced paw oedema in wister male rats. *Afric. J. Biotech*. 2007;6(10):1209-1211.
37. Adeyemi O, Okpo SO, Ogunti OO. Analgesic and anti-inflammatory effects of the aqueous extract of leaves of *Pearsea Americana mill (lauraceae)*. *Fitoterapia*. 2002;73:375-380.
38. Haruna B, Cyril OU. Phytochemical investigation and anti-inflammatory property of ethanol-water extract of the root of *Anthocleista djalonensis*. *Cher. (Gentianaceae)*. *Afric. J. Biotech*. 2011; 10(34):6598-6600.
39. Pagana KD, Pagana JJ. *Measuring white blood cells. Mosby's manual of diagnostic and lab. Tests*. st. Loris; 1978.
40. Larson A, Haux C, Sjobak M. Fish physiology and metal pollution results and experience from laboratory and food studies. *E cotox. Enrinom. Safety*. 1985;9: 250-281.
41. Vane JR. Inhibition of prostaglandin synthesis as a mechanism of action for aspirin-like drugs. *Nature (New Biol)*. 1971;132:232-235.
42. Vineger R, Schreiber W, Hope R. Biphasic development of carrageenan in rats. *J. Pharmacol. Exp. Ther*. 1969;166:96-103
43. Lawrence DR, Benneth PN, Brown MJ. *Clinical pharmacology*. Churchill Living Stone. Edinburgh London. 8th edition; 1997.

© 2016 Odin et al.; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history:

The peer review history for this paper can be accessed here:

<http://sciencedomain.org/review-history/16221>