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# Growth and Quality Profiency of Onion (Allium cepa L.) by Application of Biofertilizer, GA<sub>3</sub> and Humic Acid

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### Authors' contributions

This work was carried out in collaboration among all authors. All authors read and approved the final manuscript.

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#### ABSTRACT

During the Rabi season in 2022-2023, a comprehensive trial was carried out at the Department of Horticulture, Jawaharlal Nehru Krishi Vishwa Vidyalaya. Employing a Randomized Completely Block Design (RCBD), the research assessed twelve different interventions, consisting of biofertilizers,  $GA_3$ , humic acid and a reference group. Notably, the combined application of Azotobacter, Azospirillum,  $GA_3$  and humic acid (T10) showcased substantial improvements in several growth parameters. These encompassed plant height (62.23 cm), number of leaves (10.18), length of leaves (54.08 cm), leaf area (624.65 cm<sup>2</sup>), leaf area index (4.16) and leaf diameter (8.14

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mm). Additionally, this treatment recorded the highest Total Soluble Solids (TSS) at 12.37 °Brix. Moreover, the inclusion of sulphur and humic acid significantly affected the pyruvic acid content, with T10 recorded the highest pyruvate values (5.28 µmol/g). Conversely, the reference group (T12) only recorded slight improvements, emphasizing the noteworthy impact of the interventions on both the growth and qualitative characteristics of the onions.

Keywords: Biofertilizers; GA<sub>3</sub> and humic acid; onion growth parameters; total soluble solids (TSS); pyruvic acid content.

### 1. INTRODUCTION

The onion, referred to as the "Ruler of the Kitchen," is a biennial herb that has become a fundamental part of cooking customs worldwide. Originating from Central Asia and the Mediterranean area, it has transformed into a vital seasoning crop. It is widely farmed in different parts of India throughout both the kharif and rabi seasons, indicating its importance in the farming landscape.

Onions (Allium cepa L.), which are highly prized for their high nutritional content, are widely produced for vear-round eating [1,2]. Biofertilizers. which are microorganism inoculants used in seed inoculation, convert inaccessible nutrients into accessible forms, hence improving crop growth [3,4]. According to research by Wu et al. and Kumar et al. [5,6], these biofertilizers boost both the quality and quantity of onions.

The use of bio-fertilizers, like Azotobacter and Azospirillum, provides a cost-effective and effective approach to enhance crop growth by boosting nitrogen and phosphorus absorption, thereby significantly increasing onion output. Furthermore, gibberellic acid (GA3), a crucial plant growth controller, plays a key role in the onion's development cycle. When used skilfully, GA<sub>3</sub> promotes stem elongation, internode expansion and cell division, contributing to the development of larger bulbs. Humic acid, an organic substance, also plays a major part by enhancing soil structure, nutrient assimilation, root growth and water retention, all of which collectively lead to improved onion growth and better bulb quality.

The strategic and skilful use of these natural components is vital for maximizing onion farming results. By using bio-fertilizers and growth controllers, not only can quality be enhanced, but sustainable agricultural practices can also be promoted by reducing reliance on synthetic inputs. By harnessing the potential of these natural growth boosters, farmers can guarantee the sustainable and productive cultivation of onions, satisfying the growing demand for this versatile herb in various cooking customs.

### 2. MATERIALS AND METHODS

The research was conducted at Jawaharlal Nehru Krishi Vishwa Vidyalaya, JNKVV Jabalpur during Rabi season of 2022-23. The focus of the study was on the Bhima Shakti variety of onion. To conduct the study, a Randomized Complete Block Design was used with three replications and 12 treatments. The onions were grown in plots measuring 3x2 meters with 1-meter gaps between replications. The distance between the onions was 10 cm, while the distance between the rows was 15 cm. The objective of the study was to assess the effectiveness of a mixture of Azotobacter and Azospirillum, combined with 5 kg/ha of Farmyard Manure (FYM) during transplantation, in enhancing nitrogen fixation and nutrient availability. Furthermore, 2 kg/ha of humic acid and 100 ppm of Gibberellic acid (GA<sub>3</sub>) were applied to the leaves 60 days after transplantation to improve nutrient absorption, promote growth and increase yield. The impact of these interventions was evaluated 90 days after transplantation.

T1 received RDF (NPKS:100:50:50:40 Kg/ha), T2 received RDF + Azotobacter (5kg/ha), T3 received RDF + Azospirillum (5kg/ha), T4 received RDF + GA<sub>3</sub> (100 ppm), T5 received RDF + Humic acid (2kg/ha), T6 received RDF + Azotobacter (5kg/ha) + Azospirillum (5kg/ha), T7 received RDF + GA<sub>3</sub> (100 ppm) + Azotobacter (5kg/ha), T8 received RDF + GA<sub>3</sub> (100 ppm) + Azospirillum (5kg/ha), T9 received RDF + GA<sub>3</sub> (100 ppm) + Humic acid (2kg/ha), T10 received RDF + GA<sub>3</sub> (100 ppm) + Azotobacter (5kg/ha) + Azospirillum (5kg/ha), + Humic acid (2kg/ha), T11 received RDF + Humic acid + Azotobacter (5kg/ha) + Azospirillum (5kg/ha), and T12 served as the control group (Fig. 1).

Treatments No.	Treatment Details				
T <sub>1</sub>	RDF (NPKS:100:50:50:40 Kg/ha)				
T <sub>2</sub>	RDF + Azotobacter (5kg/ha)				
T <sub>3</sub>	RDF + Azospirillum (5kg/ha)				
Τ <sub>4</sub>	RDF + GA <sub>3</sub> (100 ppm)				
T₅	RDF + Humic acid (2kg/ha)				
T <sub>6</sub>	RDF + Azotobacter + Azospirillum				
T <sub>7</sub>	RDF + GA <sub>3</sub> + Azotobacter				
Τ <sub>8</sub>	RDF + GA <sub>3</sub> + Azospirillum				
T9	RDF + GA <sub>3</sub> + Humic acid				
T <sub>10</sub>	RDF + GA <sub>3</sub> + Azotobacter + Azospirillum + Humic acid				
T <sub>11</sub>	RDF + Humic acid + Azotobacter + Azospirillum				
T <sub>12</sub>	Control				

Table 1. Overview of Treatments

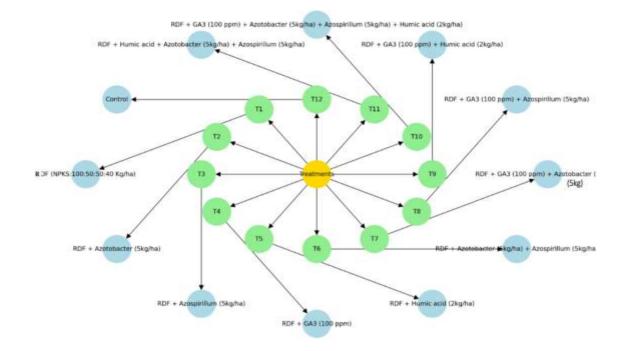


Fig. 1. Treatments details

The observations for the following parameters were recorded using standard methodology:

Leaf Area per plant (cm<sup>2</sup>): Leaf area measure by leaf area meter.

Leaf Area Index (LAI): LAI was measured with mainly based on leaf area divided by ground area occupied by plant.

Leaf area index is the ratio of leaf area to ground area.

$$LAI = \frac{Leaf Area}{Ground Area}$$

**TSS estimation:** TSS or Total Soluble Solids, is quantified using a refractometer. This device is employed to measure the concentration of dissolved solids, often in the form of sugars, in a liquid sample, providing a valuable metric for assessing the sweetness and quality of substances like fruit juices and other beverages.

**Pyruvic acid estimation:** To determine the concentration of pyruvic acid, we mixed 0.5 ml of onion extract with 1.5 ml of 5% TCA and 18 ml of distilled water. After that, we combined 1ml of this mixture with 1ml each of 2,4-DNPH and distilled water, and let it incubate at 37°C for 10 minutes. Then, we added 5 ml of 0.6 N NaOH and measured the absorbance at 420

nanometres using a Spectrophotometer to establish a standard curve. The concentration of pyruvic acid is a crucial indicator of onion pungency, determined by the breakdown of taste precursors. This helps in evaluating the flavour and sensory characteristics of the onions being studied.

**Pungency range:** The onions are divided into three categories based on their pungency: low, which is  $(0-3 \ \mu mol \ pyruvic \ acid/g)$ ; medium, which is  $(3-7 \ \mu mol \ pyruvic \ acid/g)$ ; and high, which is (greater than 7  $\ \mu mol \ pyruvic \ acid/g)$ . According to Bhima Shakti data, it can be categorized as medium/sweet pungent  $(3-7 \ \mu mol)$  [7].

### 3. RESULTS AND DISCUSSION

After 60 days of planting, treatment T10, which consisted of a comprehensive combination of Recommended Dose of Fertilizer (RDF). Gibberellic Acid (GA<sub>3</sub>), Azotobacter, Azospirillum and Humic acid, resulted in the tallest onion plants measuring 50.31 cm. This height was closely matched by T11 (49.68 cm), which used a similar mixture excluding GA<sub>3</sub>. In contrast, the control group T12 had the shortest height at 41.92 cm. By 90 days after planting, T10 continued to lead with a height of 62.23 cm, closely followed by T11 at 61.47 cm, while T12 registered a height of 51.88 cm. The significant growth response to the combination treatments of T10 indicates its effectiveness in promoting cell elongation and root expansion, which is consistent with the findings of Sajid et al. (2012) and Mohammed and Ibraheem [8,9].

In terms of foliar characteristics. T10 exhibited superior leaf count, length, area and diameter. At 60 days after planting, T10 had the highest leaf count (7.15) and length (44.91 cm), followed closely by T11 with 6.95 leaves and a considerable leaf area. By 90 days after planting, T10 maintained its lead with a leaf count of 10.18 and a length of 54.08 cm, which were nearly equivalent to T11's 9.45 leaves and significant leaf area of 596.03 cm<sup>2</sup>. On the other hand, T12 consistently showed the lowest values in all these attributes. When it comes to leaf area. T10 had the largest measurements at both 60 days after planting (320.69 cm<sup>2</sup>) and 90 days after planting (624.65 cm<sup>2</sup>), while T12 consistently recorded the smallest leaf area (241.15 cm<sup>2</sup> at 60 days after planting and 360.75 cm<sup>2</sup> at 90 days after planting). These trends highlight the positive influence of the treatment components on foliar

development, which aligns with the findings of Singh et al., Gowda et al. and Kumar et al. [10,11,6].

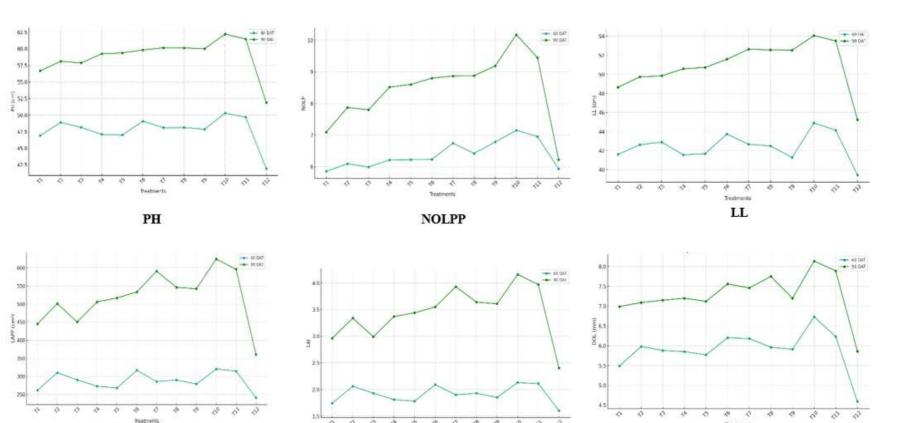
The Leaf Area Index (LAI) showed similar patterns, with T10 demonstrating the highest values at both 60 and 90 days after transplanting (DAT), recording 2.13 and 4.16 respectively. T11 closely paralleled these figures with an LAI of 2.11 and 3.97, while T12 consistently had the lowest values (1.60 and 2.40). In terms of leaf diameter, T10 led with the largest measurements at 60 DAT (6.73 mm) and 90 DAT (8.14 mm), similar to T11's figures, while T12 consistently had the smallest diameters (Table 2 and Fig. 2). These results emphasize the significant impact of the treatment components on plant growth and their importance in assessing LAI, which supports the observations of Calvo et al., Singh et al. [12,10].

Quality parameters: The impact of different treatments, namely Azotobacter, Azospirillum, GA<sub>3</sub> and humic acid, on the Total Soluble Solids (TSS) content in onions is emphasized by this research. The results showed that Treatment T10, which consisted of RDF, GA<sub>3</sub>, Azotobacter, Azospirillum and Humic acid, recorded the highest TSS value at 12.37 °Brix. Treatment T11, which included RDF, Humic acid, Azotobacter and Azospirillum, closely followed with a TSS value of 12.08 °Brix. Conversely, Treatment T12 (Control) had the lowest TSS value at 9.00 °Brix. treatments significantly These enhanced photosynthesis, optimized nutrient utilization and stimulated sugar synthesis, leading to an increase in the TSS content of Treatment T10. This finding supports previous research [13,14], highlighting the effectiveness of these treatments in increasing TSS levels in onions.

Furthermore, the study provides compelling evidence of the significant effect of these treatments on the pyruvic acid content in onion plants. Treatment T10 once again had the highest pyruvic acid content at 5.28 µmol/gm, followed closely by Treatments T11 and T9, which recorded 5.25 µmol/g and 5.24 µmol/g respectively and were statistically equivalent. In contrast, Treatment T12 (Control) had the lowest pyruvic acid content at 3.6 µmol/g. The inclusion of sulphur and humic acid played a crucial role in enhancing pyruvic acid metabolism, leading to increased energy production and improved cellular metabolic efficiency. This study aligns with previous findings by Denre et al. [15] and highlights potential opportunities for enhancing

S. No.	Treatments	Plant height (cm)		Number of leaves/plant		Leaf length (cm)		Leaf area per plant (cm²)		Leaf area index		Diameter of leaf (mm)	
		60	90	60	90	60	90	60	90	60	90	60	90
		DAT	DAT	DAT	DAT	DAT	DAT	DAT	DAT	DAT	DAT	DAT	DAT
T1	RDF (NPKS: 100:50:50:40 Kg/ha)	46.91	56.68	5.85	7.09	41.61	48.66	261.98	445.14	1.74	2.96	5.49	6.99
T2	RDF + Azotobacter (5kg/ha)	48.90	58.11	6.09	7.87	42.63	49.74	310.37	501.26	2.06	3.34	5.98	7.09
Т3	RDF + Azospirillum (5kg/ha)	48.15	57.88	5.99	7.80	42.9	49.85	290.67	450.40	1.93	2.99	5.88	7.15
T4	RDF + GA <sub>3</sub> (100 ppm)	47.09	59.25	6.21	8.52	41.57	50.59	272.99	505.83	1.81	3.37	5.85	7.20
T5	RDF + Humic acid (2kg/ha)	47.00	59.39	6.22	8.60	41.69	50.73	267.79	516.74	1.78	3.44	5.77	7.12
T6	RDF + Azotobacter + Azospirillum	49.09	59.84	6.23	8.80	43.74	51.59	317.14	533.16	2.09	3.55	6.20	7.56
T7	RDF + GA <sub>3</sub> + Azotobacter	48.11	60.15	6.74	8.87	42.67	52.65	285.77	591.05	1.90	3.93	6.18	7.46
T8	RDF + GA <sub>3</sub> + Azospirillum	48.13	60.13	6.42	8.88	42.50	52.58	290.22	546.06	1.93	3.64	5.96	7.75
Т9	RDF + GA₃ + Humic acid	47.88	60.01	6.78	9.19	41.29	52.54	279.08	542.32	1.85	3.61	5.91	7.20
T10	RDF + GA₃ + Azotobacter + Azospirillum + Humic acid	50.31	62.23	7.15	10.18	44.91	54.08	320.69	624.65	2.13	4.16	6.73	8.14
T11	RDF + Humic acid + Azotobacter + Azospirillum	49.68	61.47	6.95	9.45	44.15	53.52	314.74	596.03	2.11	3.97	6.23	7.89
T12	Control	41.92	51.88	5.93	6.22	39.45	45.25	241.15	360.75	1.60	2.40	4.59	5.86
	Sem±		0.33		0.34		0.27		16.09		0.10		0.31
	CD at 5%		0.99		0.99		0.81		47.18		0.31		0.92

## Table 2. Response of biofertilizer, GA<sub>3</sub> and humic acid on growth parameters



Treatments

DOL

Fig. 2. Comparison of PH (plant height), NOLPP (number of leaves per plant), LL (leaf length), LAPP (leaf area per plant), LAI (leaf area index), DOL (diameter of leaf) at 60 and 90 DAT (Days after Transplanting)

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S. No.	Treatment	TSS (°Brix)	Pyruvic acid (µmol/g)	Pungency
T1	RDF (NPKS: 100:50:50:40 Kg/ha)	10.04	4.12	Medium pungent
T2	RDF + Azotobacter (5kg/ha)	10.76	4.09	medium pungent
Т3	RDF + Azospirillum (5kg/ha)	10.34	4.10	medium pungent
T4	RDF + GA <sub>3</sub> (100 ppm)	10.10	4.22	medium pungent
T5	RDF + Humic acid (2kg/ha)	10.17	5.12	medium pungent
Т6	RDF + Azotobacter + Azospirillum	11.23	4.23	medium pungent
T7	RDF + GA <sub>3</sub> + Azotobacter	11.03	4.13	medium pungent
T8	RDF + GA <sub>3</sub> + Azospirillum	11.00	4.20	medium pungent
Т9	RDF + GA <sub>3</sub> + Humic acid	10.64	5.24	medium pungent
T10	RDF + GA <sub>3</sub> + Azotobacter + Azospirillum + Humic acid	12.37	5.28	medium pungent
T11	RDF + Humic acid + Azotobacter + Azospirillum	12.08	5.25	medium pungent
T12	Control	9.00	3.06	medium pungent
	Sem±	0.32	0.10	
	CD at 5%	0.96	0.30	

# Table 3. Response of biofertilizer, GA<sub>3</sub> and humic acid on quality parameters

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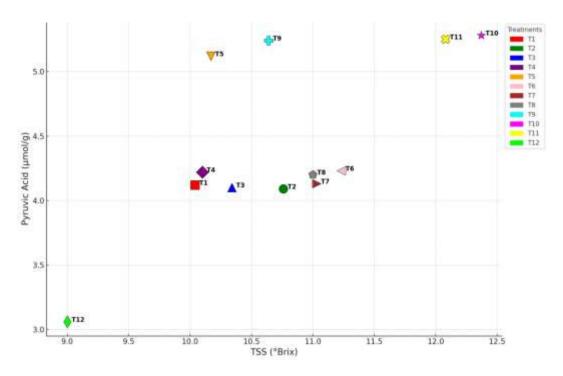


Fig. 3. Custom visualization of TSS and pyruvic acid with annotations

the nutritional characteristics of onions. Based on the data, Bhima Shakti onions are classified as having medium/sweet pungency, with a pyruvic acid content ranging from 3-7  $\mu$ mol/g (Table 3 and Fig. 3) [16].

### 4. CONCLUSION

To achieve exceptional growth and unmatched quality in cultivating the Bhima Shakti onion variety by judicial application of different treatment approaches. Particularly the combined use of Azotobacter (5 kg), Azospirillum (5 kg), GA<sub>3</sub> (100 ppm) and humic acid (2 kg) representing the treatment T10 demonstrated an impressive growth include reaching an ideal plant height of 62.23 cm, maximizing the number of leaves per plant to 10.18, maintaining an average leaf length of 54.08 cm, achieving a considerable leaf area of 624.65 cm<sup>2</sup> per plant, promoting a strong Leaf Area Index (LAI) of 4.16 and securing a sufficient leaf diameter of 8.14 mm. At the same time, it is essential to address quality indicators such as Total Soluble Solids (TSS) at 12.37 °Brix and pyruvic acid concentrations at 5.28 µmol/g to ensure a harvest of exceptional quality.

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#### **COMPETING INTERESTS**

Authors have declared that no competing interests exist.

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