

Arsenic (III) Removal from Real-Life Groundwater by Adsorption on Neem Bark (*Azadirachta indica*)

Tasrina R. Choudhury¹, M. N. Amin^{2*}, S. B. Quraishi¹ and A. I. Mustafa²

¹*Analytical Chemistry Laboratory, Chemistry Division, Atomic Energy Centre, Dhaka-1000, Bangladesh.*

²*Department of Applied Chemistry and Chemical Engineering, Dhaka University, Bangladesh.*

Authors' contributions

This work was carried out in collaboration between all authors. Author MNA designed the study. Authors MNA, AIM and SBQ supervised the work study. Author TRC carried out the most experimental works according to the plan and prepared the manuscript. Author MNA wrote the final draft of the manuscript. All authors read and approved the final manuscript.

Original Research Article

Received 4th December 2012

Accepted 8th April 2014

Published 16th June 2014

ABSTRACT

The adsorption characteristics of arsenic on neem bark, a low-cost natural adsorbent, were studied in the laboratory scale using real-life sample. Neem bark has tremendous potential as a remediation material for the removal of arsenic from groundwater. The present work investigates the possibility of the use of neem bark adsorption technology without any pretreatment in the removal of arsenic from aqueous media. Various conditions that affect the adsorption/desorption of arsenic are investigated. Adsorption column methods show the maximum removal of As (III) under the following conditions: initial arsenic concentration, 100 $\mu\text{g/L}$; neem bark amount, 5 g; average particle size, 0.595 mm; treatment flow rate, 1.67 mL/min; and pH, 6.9; respectively. The desorption efficiencies with 1M of HCl after the treatment of groundwater were in the range of 79%. The present study might provide new avenues to achieve the arsenic concentration for drinking water recommended by Bangladesh and the World Health Organization (WHO).

*Corresponding author: Email: namin@du.ac.bd;

Keywords: Arsenic (III); adsorption; neem bark; aqueous solution.

1. INTRODUCTION

Arsenic (As) is mostly found in the earth's core and in clay- and sulphide-rich portions of the earth's crust [1]. Being a metalloid in group 15 on the periodic table (along with antimony, bismuth, nitrogen and phosphorus), arsenic is well known for its chronic toxicity, particularly when exposure occurs over prolonged periods [2]. Arsenic exposure via drinking-water is related to lung, kidney, bladder and skin cancer. For example, drinking-water arsenic concentrations in excess of 50 $\mu\text{g/L}$ have been associated with increased risks of cancer in the bladder and lung, whilst drinking-water arsenic levels even below 50 $\mu\text{g/L}$ have been associated with precursors of skin cancer [3]. Therefore, the presence of arsenic in water supply poses a serious risk to human health. Surface and ground waters in many parts of the world have been found to naturally contain As concentrations that make these waters unsuitable for human use. Significant concentrations of As have been reported in various countries such as Bangladesh, Chile, USA, China, and India. In Bangladesh, for example, about 100 million people currently drink water with As concentrations up to 100 times the World Health Organisation (WHO) drinking water guideline, which is 10 $\mu\text{g/L}$ [4].

To remove As from potential drinking water sources, a variety of conventional and non-conventional technologies have been studied, and these technologies have been reviewed by several authors [4]. However, it is known that conventional engineered treatment technologies are costly and create problems of sludge generation and disposal [5–7]. In addition, these systems often become sources of As-rich effluents and are typically located in remote isolated areas (such as mining sites), thus precluding the transportation of the effluents to large centralised treatment facilities. As such, to prevent arsenic (III) pollution of water courses, it is essential to find onsite, decentralised treatment systems that are robust and have low maintenance requirements and operating costs.

Many technologies, including coprecipitation with iron or alum, adsorption onto coagulated floc, ion-exchange resin, reverse osmosis, and membrane techniques, have been used to remove As from aqueous solution [8–14]. In these methods, however, the adsorption techniques are simple and convenient and have the potential for regeneration and sludge-free operation. So far, various adsorbents for arsenic removal have been developed that include such materials as metal-loaded coral limestone [15–16], hematite and feldspar [17], activated carbon [18–19], activated alumina [20–21], and hydrous zirconium oxide [22]. However, most of these adsorbents entail several problems in terms of efficiency and cost. The present work was performed to evaluate the use of neem bark without any pretreatment as an alternate adsorbent for removing arsenite from aqueous mediums. Moreover, the neem bark was applied to the removal of arsenic from Bangladeshi As-contaminated drinking water samples in a single-step column operation.

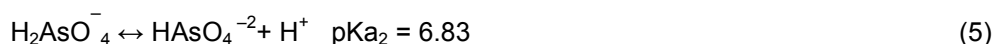
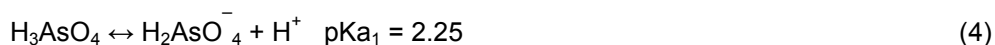
1.1 Arsenic Chemistry

Arsenic is a highly reactive metalloid that can be found in oxidation states -3, 0, +3 and +5. In natural waters, arsenic occurs as arsenite (AsO_3^{-3}) and arsenate (AsO_4^{-3}), referred to as As (III) and As (V), respectively. As (III) mostly exists in reducing groundwaters and hydrothermal waters, while As (V) is more often present in surface waters and oxidising groundwaters [23]. The main factors that control arsenic speciation are the oxidation state and pH. As (III) commonly hydrates to arsenious acid; therefore its chemistry depends

strongly on pH. The predominant As (III) species is arsenious acid, H_3AsO_3 , due to the high value of pK_{a1} [24] under the condition of 25°C and 1 bar pressure. The reactions of the dissociation of arsenious acid and its respective anions are shown below, along with the associated pK_{as} values (where $pK_a = -\log(K_a)$, and $K_a =$ equilibrium constant of the reaction):



As (V) commonly hydrates to arsenic acid, and its chemistry also depends on pH. The most common species are $H_2AsO_4^-$ and $HAsO_4^{-2}$, due to the low pK_{a1} value [24]. The reactions of dissociation of arsenic acid and its respective anions are shown below along with the associated pK_a values:



The toxicity of arsenic depends on its speciation; for example, arsenite is significantly more toxic than arsenate [25]. It is typically more difficult to remove arsenite than arsenate from contaminated water; this is because in natural waters, under normal pH conditions (6-9), arsenite is mostly found as an uncharged species (H_3AsO_3), and negatively charged species ($H_2AsO_3^-$, $HAsO_3^{-2}$ and AsO_3^{-3}) are found only at high pH (>9). On the other hand, arsenate is commonly found as negatively charged species ($H_2AsO_4^{-2}$ and $HAsO_4^{-2}$); which are easier to remove than are uncharged species, because the charges of the As (V) oxyanions allow them to be removed by sorption, anion exchange or precipitation/coprecipitation [23].

2. MATERIALS AND METHODS

2.1 Reagent

All reagents used throughout this work were of analytical-grade purity. NaOH, KOH, KI, HCl, HNO_3 , and H_2SO_4 were obtained from E. Merck, Germany. Individual standard solution of As (III) was supplied by Varian Inc, USA with highest purity level (99.98%). Dilute standard solutions were prepared daily before use. Pure water was purified with an ultrapure water system (Barnsted E-Pure, USA) resulting in a resistivity of >18 MΩ cm.

2.2 Preparation of Adsorbent

The neem bark used in the present work was obtained from Bangladesh. The chemical composition of neem bark has been reported as 3.43 wt % protein, 0.68 wt % Alkaloids, 4.16 wt % Mineral. The collected materials were washed with pure water several times to remove dust and fines. The washing process was repeated until the color of the wash water was transparent. The washed materials were then dried in a hot-air oven at 60°C for 24 h. The dried material was sieved into the following five size fractions: 0.105mm, 0.25mm, 0.595mm,

1.41mm, and 2.3 mm. The materials were used for the removal of arsenic without further physical or chemical treatment.

2.3 Adsorption and Analytical Procedures

Neem bark was added to the treatment glass columns (2 x 30 cm). The adsorption experiments were carried out in columns that were equipped with a stopper for controlling the column eluate flow rate (treatment rate). Adsorption factors including the amount of duckweed (1-5 g), average particle size (0.105-2.3mm), treatment flow rate (3.3-0.41 mL/min), initial sample concentration (50-500 µg/L), and pH (2.3-12) were evaluated. After the pH had been adjusted to the desired value with HCl and NaOH solutions, the sample solution (100 mL) was passed through the adsorption column at a given flow rate. The treatment flow rates of 0.41, 0.55, 0.83, 1.67 and 3.3 mL/min correspond to 5 g of neem bark. The packing density of the treatment column was 0.32 g/cm³. A small piece of tissue paper was inserted into the bottom of the column to prevent the loss of neem bark during the treatment. The flow rate was kept constant by controlling the stopper valve. The removal treatment was performed at ambient temperature. The number of experiments for the removal of As was greater than five. Analyses were performed by Varian Atomic Absorption Spectrometer (Model SpectrAA 240). The instrument is equipped with a Hydride generation system (Model VGA-77) and controlled with software Version 5.01. Samples were analyzed with HG-AAS using three-point calibration.

In the analysis time, acid container contains 5M HCl and reductant container contain 0.6 % Sodium borohydride (NaBH₄) in 0.5 % (w/v) sodium hydroxide.

The removal (adsorption) efficiency was calculated using the equation,

$$\text{Removal (adsorption) efficiency} = \frac{C_o - C_t}{C_o} \times 100$$

Where C_o and C_t are the concentration of As in the sample solution before and after treatment, respectively.

3. RESULTS AND DISCUSSION

First, the performances of eight adsorbents (Coconut shell, neem bark, orange peel, nut shell, flax, saw dust, wheat stalk, newspaper) were evaluated for the removal of As (III) from aqueous solutions. The removal efficiencies with coconut shell, neem bark, orange peel, nut shell, flax, saw dust, wheat stalk and newspaper were 18.79%, 19.23%, 3.8%, 14.26%, 12.89%, 14.2%, 12.3%, 32.3%, respectively. Although the adsorption capacity of news paper was highest, neem bark was taken as adsorbent because the main component of neem bark is carbon, neem bark has the potential to be used as an adsorbent. It was chosen for use as an adsorbent material because of its granular structure, insolubility in water, chemical stability, and high mechanical strength. Neem bark was selected as a suitable adsorbent. Of the adsorption neem bark gives the highest desorption and regeneration. Apart from this neem bark is widely available throughout the country.

3.1 Effect of Adsorbent Amount

The effect of the amount of neem bark on the removal of As (III) was investigated. The results are presented in Fig. 1. The removal efficiencies of As (III) increased gradually with increasing amount of neem bark. It is readily understood that the adsorption capacity of neem bark depends on the surface activity, that is, the specific surface area available for As-surface interactions that is accessible to the As. Hence, increasing the amount of neem bark will increase removal capacity of As. Until now, various kinds of adsorbents have been studied for the removal of arsenic. The maximum As (III) removal efficiency achieved was 99% for iron oxide-coated sand at an adsorbent dose of 20 g/L with an initial As concentration 100 $\mu\text{g/L}$ in batch studies [26]. The removal efficiencies for As(III) from an aqueous solution (100 $\mu\text{g/L}$, 100 mL) by 0.1 g of modified fungal biomass were 75%, respectively, after a 12-h batch treatment [27]. When an aqueous As (III) solution of 10 mg/L concentration was stirred in the presence of both 1.0 g/L TiO_2 and 1.0 g/L activated alumina under sunlight irradiation, the arsenic removal increased with time and reached 89% after 24 h [28]. Although the removal efficiencies in the proposed system were similar to those obtained with the other adsorbents, the treatment time was very short because of the flow method (column system).

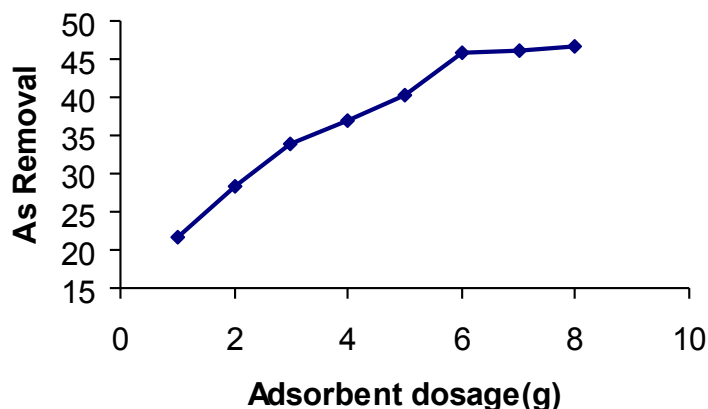


Fig. 1. Effect of adsorbent amount on the removal of As (III) by adsorption onto neem bark (initial As concentration, 100 $\mu\text{g/L}$; pH 2.36; Treatment flow rate, 1.67mL/min)

3.2 Effect of Flow Rate

Flow rate is one of the effective factors in column adsorption process. The effect of the treatment flow rate on the removal of As (III) is illustrated in Fig. 2. The removal efficiency for As (III) increased gradually with decreasing treatment flow rate. This is because, when the flow rate was slow, As (III) in the sample solution got more contact time with the active surface of the adsorbents.

3.3 Effect of Initial Concentration

The removal efficiency is highly dependent on the initial concentrations of As (III) in the sample solution. The effect of the initial sample concentration on the removal of arsenic with neem bark was investigated. The initial concentration was evaluated in the range of 50-500

$\mu\text{g/L}$, and the results are illustrated in Fig. 3. The removal efficiency decreased with increasing adsorbate concentration in the solutions. It is because, at low concentration, most of As (III) in the sample solution gets contact with active sites. With increasing arsenic concentration in solution, active sites on the adsorbents were not increased. Therefore arsenic removal efficiencies were decreased.

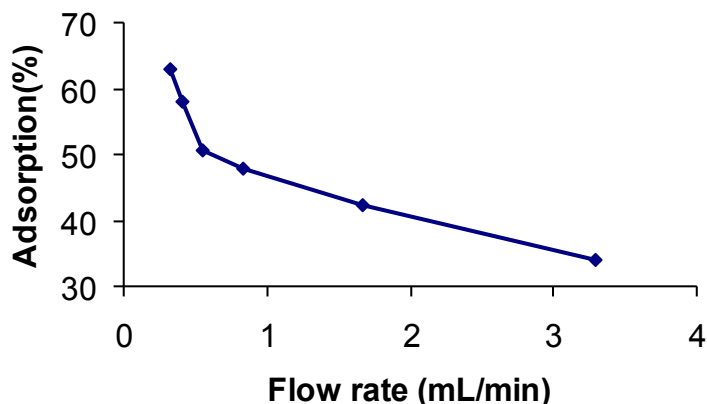


Fig. 2. Effect of treatment flow rate on the removal of As (III) by adsorption onto neem bark (initial As concentration, $100 \mu\text{g/L}$; pH 2.36; Amount of adsorbent, 5g)

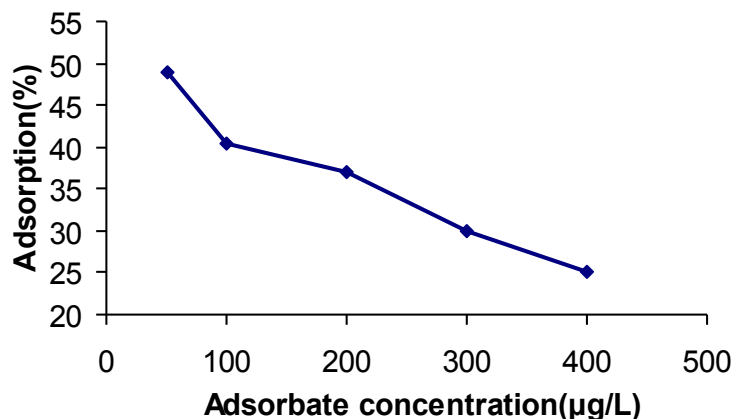


Fig. 3. Effect of initial adsorbate concentration on the removal of As (III) by adsorption onto neem bark (pH 2.36; Amount of adsorbent, 5g; Treatment flow rate, 1.67 mL/min)

3.4 Effect of pH

pH is one of the most important parameters controlling the metal ion sorption process [29–30]. Fig. 4 depicts the effect of pH on As(III) removal with neem bark. For the removal of As (III), the efficiency curve was essentially nearly a plateau in the pH range of 4-7, and then, the efficiency tended to decrease with increasing pH. The poor As removal efficiencies at high pH can be attributed to the following factors: First, chemical species for As (III) in this pH region are oxyanions. Next, hydroxyl groups are more plentiful on the surface of neem bark with increasing pH. The maximum removal efficiency for As (III) was observed in

comparatively neutral region (at pH 7). This result should be of great advantage for the practical implementation of arsenic removal from groundwater.

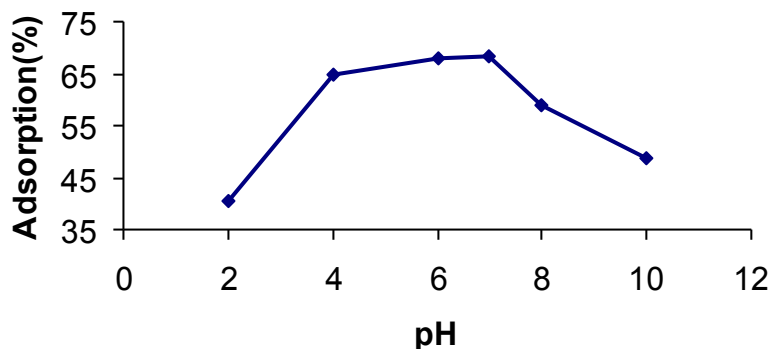


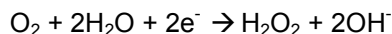
Fig. 4. Effect of pH on the removal of As (III) by adsorption onto neem bark (initial As (III) concentration, 100 µg/L; Amount of adsorbent, 5g; Treatment flow rate, 1.67mL/min)

3.5 Effect of Particle Size

Column adsorption experiments were carried out for the removal of arsenic from aqueous solution using five different particle sizes (0.105mm, 0.25mm, 0.595 mm, 1.41mm, and 2.3 mm). The results are shown in Fig. 5. With decreasing particle size, the removal increased from 68.23% to 96% for As (III). Munaf and Zein reported that, when the size of adsorbent particles increased, the adsorption of metal ions decreased [29]. Similar trends have been reported by Wong et al. [30]. These phenomena might be due to the fact that the smaller particles offer comparatively larger surface areas and greater numbers of adsorption sites.

3.6 Adsorption Mechanism

Arsenic removal by adsorption onto neem bark can be supposed to occur mainly through two routes: (i) affinity adsorption and (ii) anion exchange between the arsenic in the water and the carbon surface of the neem bark. Affinity adsorption is related to the surface behavior of neem bark, whereas anion exchange relates to the existing forms of the arsenic species. OH groups are created on the carbon surface during the activation process [31–32]. The mechanism of adsorption of metal anions onto activated carbon is generally well explained by electrochemical theory: Carbon in contact with water reduces oxygen to a hydroxyl group [33]



And thus, the carbon loses electrons and become positively charged. Electrical neutrality is maintained with hydroxyl ions, resulting in their adsorption. Although the neem bark used in the present work was not subjected to any chemical or physical activation treatment, a large number of OH groups will remain on the surface of the neem bark after the drying process. The physical adsorption and interaction between the H_3AsO_3 species and the neem bark surface might be partly responsible for the removal of As (III).

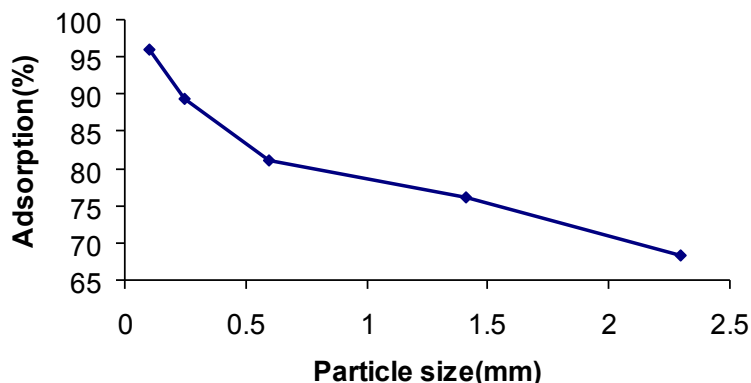


Fig. 5. Effect of particle size on the removal of As (III) by adsorption onto neem bark (initial As concentration, 100 µg/L; pH 7; Amount of adsorbent, 5g; Treatment flow rate, 1.67mL/min)

3.7 Desorption

Recovery of the adsorbed material and regeneration of the adsorbent are also important aspects of wastewater treatment. Attempts were made to desorb As (III) from the neem bark surface with various eluents, such as hydrochloric, sulfuric, and nitric acid solutions and base solutions containing sodium hydroxide and potassium hydroxide.

This desorption process was performed using the column method. For each experiment, 100 mL of desorption solution was added to the column and held there for a fixed period of time. After the standing time, the solution was passed through the column. The results are presented in Table 1. Although the achievement of arsenic elution using strong acidic or alkaline solutions has been reported in the literature [34], the present work showed that effective desorption was obtained with acidic solutions. These phenomena are consistent with the results observed for the effect of pH. Consequently, hydrochloric acid solution was useful for the desorption of arsenic from the surface of neem bark.

Table 1. Influence of the Eluent on the desorption of As (III)

| Desorption agent | Desorption (%) |
|-------------------------------------|----------------|
| NaOH (1M) | 38.0 |
| KOH(1M) | 42 |
| HCl(1M) | 79 |
| H ₂ SO ₄ (1M) | 48.08 |
| HNO ₃ (1M) | 51.63 |

Adsorption process: initial As(III) concentration, 100 µg/L; neem bark 5g; average particle size, 0.595 mm; treatment flow rate, 1.67mL/min.; Volume of desorption agent, 100mL.

3.8 Application of the Developed Treatment System

The utility of the neem bark was evaluated for the treatment of As contaminated Bangladeshi groundwater samples. The concentrations of total arsenic in the samples was 233.03 µg/L. It has been reported that the total arsenic concentration in the tubewell water is in the range

0.25-1 mg/L, with 60-90% of the arsenic present as As (III) species [35]. Because the pH of these groundwater samples was around 7, the arsenic species might be H_3AsO_3 for As (III) [36-38]. The treatment results are presented in Table 2. Although 10 g (5 g + 5 g) of adsorbent was applied in the treatment, the concentration of arsenic in the treated sample water could be lowered to 60 $\mu\text{g/L}$. The desorption efficiencies with 100 mL of 1 M HCl was 89.63%. From the present results, the arsenic was successfully removed from practical As-contaminated groundwater, and the adsorbed As could be recovered from the surface of neem bark.

Table 2. Removal and desorption of As (III) from the contaminated groundwater of Bangladesh

| | Sample |
|--|---------------|
| pH | 7.5 |
| initial As (III) conc. ($\mu\text{g/L}$) | 233.03 |
| final As (III) conc. ($\mu\text{g/L}$) | 60 |
| removal ^a (%) | 74.25 |
| desorption ^b (%) | 89.63 |

^aRemoval: neem bark 10 g, treatment flow rate 1.67 mL/min, average particle size 0.595 mm ^bDesorption: 1M HCl, 100 mL, flow rate 1.67 mL/min.

4. CONCLUSION

The presence of arsenic in groundwater has been recognized as a major problem for Bangladesh. Due to its high toxicity, even in low concentrations, it is a threat to human health. There are many techniques, which can be effectively applied to remove arsenic from water streams. Moreover, some of the existing techniques are costly and they are not economically applicable in small community systems. Therefore, there is a need for developing cheap efficient methods for the removal of arsenic from drinking water. The proposed column treatment systems are appropriate and suitable home made approaches to arsenic removal in local areas, because of their simplicity and easy operation and handling. The present method is effective for a wide range of concentrations (i.e., 10-500 $\mu\text{g/L}$), which were quite similar to those observed in contaminated Bangladeshi groundwater. No secondary-pollution problem will occur, because desorption of the arsenic is possible. Direct removal of arsenic (III) can be achieved without first oxidizing arsenite to arsenate, whereas the traditional methods require the oxidation process. Based on the results of this research, neem bark can be considered as low cost, effective, available and natural adsorbent for removing arsenic from ground water.

ACKNOWLEDGEMENTS

The authors are grateful to the chairman of the Department of Applied Chemistry and Chemical Engineering, University of Dhaka, Bangladesh for providing the laboratory facilities during the progress of this work. The authors are also delighted to express their thanks to the authority of Bangladesh Atomic Energy Commission, Dhaka, Bangladesh for providing laboratory facilities to analyze the samples using conventional technique.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

1. Henken KR. Introduction. In: Henken KR, (Ed.), *Arsenic Environmental Chemistry, Health Threats and Waste Treatment*. John Wiley & Sons Ltd. Chichester. 2009;1–7.
2. Katherine LA, Tim DF, Guangzhi S. Removal processes for arsenic in constructed wetlands. *Chemosphere*. 2011;84:1032–1043.
3. IPCS. *Arsenic and Arsenic Compounds*. World Health Organisation. International Programme on Chemical Safety (Environmental Health Criteria 224), Geneva; 2001.
4. Mohan D, Pittman JCU. Arsenic removal from water/wastewater using adsorbents—a critical review. *J Hazard Mater*. 2007;142:1–53.
5. Kosolapov DB, Kusch P, Vainshtein MB, Vatsourina AV, Wießner A, Kästner M, Müller RA. Microbial processes of heavy metal removal from carbon deficient effluents in constructed wetlands. *Eng Life Sci*. 2004;4:403–411.
6. Cohen RRH. Use of microbes for cost reduction of metal removal from metals and mining industry waste streams. *J Clean Prod*. 2006;14:1146–1157.
7. Nelson EA, Specht WL, Knox AS. Metal removal from water discharges by a constructed treatment wetland. *Eng Life Sci*. 2006;6:26–30.
8. Katsoyiannis IA, Zouboulis AL. Application of biological processes for the removal of arsenic from groundwaters. *Water Res*. 2004;38:17.
9. Kim J, Benjamin MM. Modeling a novel ion exchange process for arsenic and nitrate removal. *Water Res*. 2004;38:20–53.
10. Thakur LS, Semil P. Removal of arsenic in aqueous solution by low cost adsorbent: A short review. *International J Chem Tech Research*. 2013;5(3);1299–1308.
11. Chandra V, Park J, Young Chun Y, Lee JW, Hwang I, Kim KS. Water-dispersible magnetite-reduced graphene oxide composites for arsenic removal. *ACS, NANO*, 2010;4(7):3979–3986.
12. Jay JA, Blute NK, Hemond HF, Durant JL. Arsenic sulfides confound anion exchange resin speciation of aqueous arsenic. *Water Res*. 2004;38:1155.
13. Hege KV, Verhaege M, Verstraete W. Electro-oxidative abatement of low-salinity reverse osmosis membrane concentrates. *Water Res*. 2004;38:1550.
14. Balasubramanian N, Madhavan K. Arsenic removal from industrial effluent through electrocoagulation. *Chem Eng Technol*. 2001;24:519.
15. Maeda S, Ohki A, Saikoji S, Naka K. Iron (III) hydroxide loaded coral limestone as an adsorbent for arsenic (III) and arsenic (V). *Sep Sci Technol*. 1992;27:681.
16. Ohki A, Nakayachigo K, Naka K, Maeda S. Adsorption of inorganic and organic arsenic compounds by aluminium-loaded coral limestone. *Appl Organomet Chem*. 1996;10:747.
17. Singh DB, Prasad G, Rupainwar DC. Adsorption technique for the treatment of As(V)-rich effluents, *Colloids Surf. A: Physicochem Eng Aspects*. 1996;111:49.
18. Manjare SD, Sadique MH, Ghoshal AK. Equilibrium and kinetics studies for As(III) adsorption on activated alumina and activated carbon. *Environ Technol*. 2005;26:1403.
19. Huang CP, Fu PLK. Treatment of arsenic(V)-containing water by the activated carbon process. *J Water Pollut Control Fed*. 1984;56:233.
20. Ghosh MM, Yuan JR. Adsorption of inorganic arsenic and organoarsenicals on hydrous oxides. *Environ Prog*. 1987;6:150.

21. Hathaway SW, Rubel FJ. Removing arsenic from drinking water. *J Am Water Works Assoc.* 1987;79:61.
22. Suzuki TM, Bomani JO, Matsunaga H, Yokoyama T. Removal of As (III) and As (V) by a porous spherical resin loaded with monoclinic hydrous zirconium oxide. *Chem Lett.* 1997;11:1119.
23. Henken KR, Hutchison A. Arsenic chemistry. In: Henken KR. (Ed.), *Arsenic Environmental Chemistry, Health Threats and Waste Treatment.* John Wiley & Sons Ltd Chichester. 2009;9–68.
24. Wolthers M, Charlet L, Van DWCH, Van DLPR, Rickard D. Arsenic mobility in the ambient sulfidic environment: Sorption of arsenic (V) and arsenic (III) onto disordered mackinawite. *Geochim Cosmochim Acta.* 2005;69:3483–3492.
25. APHA, AWWA, WEF. *Standard Methods for the Examination of Water and Wastewater*, 21st ed. American Public Health Association, Washington, DC; 2005.
26. Gupta VK, Saini VK, Jain N. Adsorption of As (III) from aqueous solutions by iron oxide-coated sand. *J Colloid Interface Sci.* 2005;55:288.
27. Pokhrel D, Viraraghavan T. Arsenic removal from an aqueous solution by a modified fungal biomass. *Water Res.* 2006;40:549.
28. Nakajima T, Xu YH, Mori Y, Kishita M, Takanashi H, Maeda S, Ohki A. Combined use of photocatalyst and adsorbent for the removal of inorganic arsenic (III) and organoarsenic compounds from aqueous media. *J Hazard Mater.* 2005;120:75.
29. Munaf E, Zein R. The use of rice husk for removal of toxic metals from wastewater. *Environ Technol.* 1997;18:359.
30. Wong KK, Lee CK, Low KS, Haron MJ. Removal of Cu and Pb by tartaric acid modified rice husk from aqueous solutions. *Chemosphere.* 2003;50:23.
31. Corapcioglu MO, Huang CP. The adsorption of heavy metals onto hydrous activated carbon. *Water Res.* 1987;21:1031.
32. Manju GN, Raji C, Anirudhan TS. Evaluation of coconut husk carbon for the removal of arsenic from water. *Water Res.* 1998;32:3062.
33. Navarro P, Alguacil FJ. Adsorption of antimony and arsenic from a copper electrorefining solution onto activated carbon. *Hydrometallurgy.* 2002;66:101.
34. Lorenzen L, Deventer JSJV, Landi WM. Factors affecting the mechanism of the adsorption of arsenic species on activated carbon. *Miner Eng.* 1995;8:557.
35. Khoe GH, Emmett MT, Robins RG. Photoassisted Oxidation of Species in Solution; U.S. Patent 5. 1997;688:378.
36. Meng X, Bang S, Korfiatis GP. Effect of silicate, sulfate and carbonate on arsenic removal by ferric chloride. *Water Res.* 2000;34:1255.
37. Ghimire KN, Inoue K, Yamaguchi H, Makino K, Miyajima T. Adsorptive separation of arsenate and arsenite anions from aqueous medium by using orange waste. *Water Res.* 2003;37:4945.
38. Wickramasinghe SR, Han B, Zimbron J, Shen Z, Karim MN. Arsenic removal by coagulation and filtration: Comparison of groundwaters from the United States and Bangladesh. *Desalination.* 2004;169:231.

© 2014 Choudhury et al.; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/3.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history:

The peer review history for this paper can be accessed here:
<http://www.sciencedomain.org/review-history.php?iid=536&id=7&aid=4948>