



Hepatic Antioxidant Effect of Paroxetine in Rats Exposed to Chronic Restraint Model

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Author's contribution

This whole work was carried out by the author SMK.

Original Research Article

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ABSTRACT

Depressed mood, with its accompanying mental and physical stresses, could affect the progression and severity of several diseases e.g. hypertension, myocardial infarction, gastritis, peptic ulcer. The present animal study was done to investigate the potential antioxidant effect of paroxetine, as a selective serotonin reuptake inhibitor, to protect against chronic restraint stress-induced oxidative damage in the liver. Thirty albino rats were divided into 3 equal groups. Group 1 was control, non-stressed non-treated group. Group 2 was exposed to chronic restraint model by placing them in wire mesh cages exactly fit to their size for 6 hours daily for 21 days. Group 3 were also exposed to chronic restraint model for 21 days while they were administered by paroxetine 1 mg/kg/day ip during the restraint period. At the end of the study, liver transaminases were determined by commercial kits. The hepatic levels of glutathione peroxidase, catalase and thiobarbituric acid reactive substance were also determined by spectrophotometric methods. Glutathione repletion ability by hepatic cells with and without paroxetine treatment was also determined in all tested groups. The results showed a significant ($p < 0.05$) increase in serum levels of transaminases and liver anti-oxidant enzymes while levels of thiobarbituric acid reactive substance were significantly ($p < 0.05$) reduced in paroxetine-treated group compared with non-stressed non-treated control rats. Glutathione repletion ability was also significantly ($p < 0.05$) increased in treated group to a level comparable to the control non-stressed non-treated values.

Keywords: *Paroxetine; chronic restraint mode; rats; anti-oxidant; liver.*

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1. INTRODUCTION

Paroxetine is one of the selective serotonin reuptake inhibitors (SSRIs). It is commonly used in the treatment of patients with depressive disorders since the late 1990s [1]. It is generally, according to animal studies and human clinical trials, preferred over tricyclic antidepressants (TCAs) and monoamine oxidase inhibitors (MAOIs) due to lesser adverse effects, good compliance, and comparable efficacy with these older drug groups [2].

Chronic stress exposure is associated with neurodegeneration that leads to mood changes with an associated marked change in anti-oxidant enzymes [3]. These enzymes are known to have an important role in the prophylaxis against damage of any cell in the body by oxidative stress. Generation of free radicals and reactive oxygen species (ROS) is an ordinary result of metabolic processes that occur regularly in human body. However, in case of exposure to chronic stress, they are produced in a very high levels that lead to toxic effects on intracellular components such as DNA, proteins and lipids with a great damaging impact on the limbic system and hence on mood status [3,7]. Additionally, the signaling processes in the body would be affected in a way resulting in depressed mood and an increase in the incidence of apoptosis of cells of vital organs [4]. Similarly, liver and other body's systems are exposed to damaging effects by oxidative stress associated with depressed mood and disorders of important mediators of central nervous system [5-7].

Depressed mood in human and exposure of animals to chronic restraint model are associated with an increase in incidence of lipid peroxidation and production of ROS [8,9]. Newer generations of antidepressant drugs as venlafaxine were found to have the ability to combat the oxidative stress with their abilities to normalize or even increase the activities of potent antioxidant markers [10,11].

The aim of the present study is to investigate the possible protective effect of paroxetine, as an SSRI used in treatment of depressed mood, on the liver of albino rats exposed to chronic restraint model.

2. MATERIALS AND METHODS

2.1 Animals

Albino rats (150-200 g body weight) were purchased from the animal house (Bilharzial Center of Research, Ain Shams University, Cairo, Egypt), housed in controlled environmental conditions. They were left for one week for acclimatization. They were housed in cages with a natural light-dark cycle and fed on a standard pellet diet and water ad libitum.

2.2 Chemicals

Paroxetine HCl (Glaxo SmithKline, U.K.) was purchased as an odorless, off-white powder whose molecular weight is 329.4, N-(1-naphthyl) ethylenediamine dihydrochloride (ICN Biomedicals Inc., Egypt) and diethyl maleate (Merck Biochemicals, Egypt). Serum alanine & aspartate transaminases (ALT, AST) spectrum diagnostic kits were obtained from Biodiagnostics, Cairo, Egypt. All other chemicals were purchased from Sigma chemicals co.

2.3 Animal Grouping

Albino rats were divided into 3 groups (N= 10 rats in each group). They were classified as follows:

- Group 1: non-stressed non-treated control group which received normal diet and saline intraperitoneal [ip] as a solvent of paroxetine.
- Group 2: non-treated group but exposed to chronic restraint model and administered only by saline ip as a solvent of paroxetine.
- Group 3: Paroxetine-treated group: was administered paroxetine dissolved in saline (1mg/kg b.wt./day ip) for 21 days. This dose was selected according to many studies [12,21,22].

2.4 Ethics

All procedures were in accordance with the National Institute of Health's Guide for the Care and Use of Laboratory Animals, as well as the guidelines of the Animal Welfare Act (www.nih.gov) [22].

2.5 Justification of Sample Size

Number of rats in each group = 10 was determined by sample size determination using program of Microlab, version 2005 so as the α value is < 0.05 and β value is $> 80\%$. The 10 rats/group is the minimal number sufficient to produce these statistical values determined by the pilot study done before the full experimental research. Values will be statistically analysed using Prism version 3.0 to calculate analysis of variance [ANOVA] between different studied groups and to determine which group is responsible for the significant changes.

2.6 Chronic restraint stress procedure

Each rat of both groups 2 & 3 was placed in a wire mesh restrainer 6 hours daily for 21 days. At the end of the restraint period, rats were moved to their cages.

2.7 Experimental protocol

Rats were weighed and each one was placed in an individual cage. They were assigned to experimental or control groups (n=10 in each group). The control animals were left undisturbed during the 21 days-period, except for scheduled daily ip injection of saline simulating the test group of treated animals, in addition to cleaning, feeding and weighing procedures. At the end of the experimental period, the animals were weighted, anesthetized with urethane (1 g/Kg). Blood samples were collected from the abdominal aorta as follows: the rat was fixed on a wooden plate and the abdominal cavity was opened, then the abdominal aorta was explored at its bifurcation after gentle traction of the viscera using soft tissue. Blood was centrifuged at 5000 rpm for 10 minutes for serum separation, and kept at -80°C until measurements of bot ALT & AST [7].

Body weight of all tested rats was constant all over the study. Serum corticosterone was significantly ($p < 0.05$) increased in stressed non-treated rats of group 2 [$22.5 \pm 2.3 \mu\text{g/ml}$,

mean \pm SD] compared to control (group 1) and paroxetine-treated group (3) [0.9 ± 0.01 & 1.2 ± 0.03 $\mu\text{g/ml}$ respectively, mean \pm SD].

2.8 Biochemical Measurements

Serum levels of alanine and aspartate transaminases (ALT & AST).

They were measured using biochemistry automatic analyzer (Hitachi 7600).

Determination of Glutathione Peroxidase [GPx] Enzyme Activity.

Glutathione peroxidase (GPx) activity in the liver homogenates was measured by the method described by Rotruck, et al. [13].

Determination of Catalase Enzyme Activity Catalase (CAT) activity in the liver homogenates was assayed colorimetrically as described by Sinha, et al. [14] at 620 nm.

Hepatic lipid peroxidation Hepatic lipid peroxidation was assayed by measurement of the concentration of TBARS in nmoles/ milligram protein of hepatic tissues according to the method described by Fraga et al. [15].

The absorbance of the precipitate of hepatic tissue was measured spectrophotometrically at 532 nm using 1,1,5,5-tetraethoxypropane as standard.

Determination of Glutathione Repletion Ability by Hepatic Cells of Tested Rats.

Diethyl maleate (DEM.), a carbonyl compound that rapidly depletes cytosolic glutathione was used to compare glutathione-replenishing abilities of hepatic cells of all tested groups according to Saville [16]. The concentrations of glutathione in different samples were expressed in nmoles per milligram of protein.

2.9 Protein Determination

The protein content of the liver homogenates was measured spectrophotometrically by Bradford method [17]. Activities of both catalase and glutathione peroxidase enzymes were expressed in Unit/mg tissue protein.

2.10 Data Analysis

Results are expressed as mean \pm SD [Standard Deviation]. Statistical analysis was performed by analysis of variance followed by Tukey's post hoc using Graph Pad Prism version 3.00 for Windows 97 (Graph Pad Software, San Diego, CA, USA). Differences with $p < 0.05$ were considered to be statistically significant.

3. RESULTS

Mean \pm SD of the serum level of alanine and aspartate aminotransferase (ALT & AST) enzymes Chronic restraint of rats of group (2) significantly ($p < 0.05$) reduces Mean \pm SD of serum levels of both ALT & AST. While, paroxetine administration, in a dose of 1 mg/kg/day

ip for 21 days, significantly ($p < 0.05$) restores them to levels comparable to control non-stressed, non-treated group (1) Fig.1.

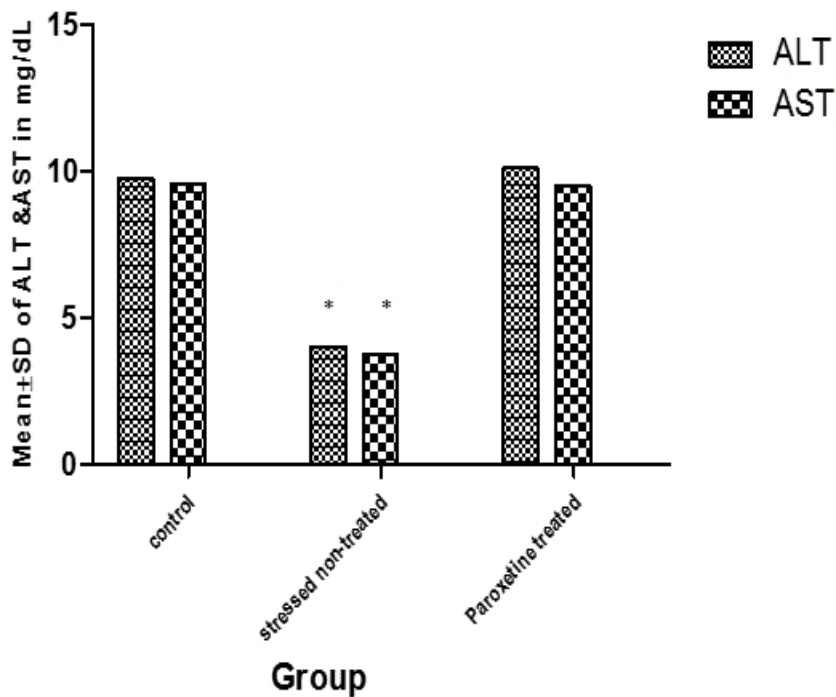


Fig. 1. Effect of treatment with paroxetine on the serum level of alanine and aspartate aminotransferase (ALT & AST) enzymes of rats compared to both groups 1 & 2 .

* Significant ($p < 0.05$) reduction in levels of both enzymes compared to control non-stressed group 1 and paroxetine-treated stressed group 3.
pearson correlation = 0.9 with perfect positive linear relationship

Table 1. Mean ± SD changes in the activities of liver CAT, GPx in rats of all tested groups expressed as unit/mg tissue protein of liver homogenates

	Control group	Stressed non-treated group	Stressed paroxetine-treated group
Glutathione peroxidase Unit ^b /mg tissue protein	12.33±1.5	0.55±0.01*	10.96±0.65**
Catalase Unit ^a /mg tissue protein	65.43±3.2	3.12±0.42*	57.65±4.5**

Table (1) ^a = Moles of hydrogen peroxide consumed per minute.

^b = μ g of glutathione consumed per minute.

* $P < 0.05$, significant reduction in activity of both enzymes in stressed non-treated group versus control group.

** $P < 0.05$, significant increase in activity of both enzymes by paroxetine administration versus stressed non-treated group.

pearson correlation = 0.9 with perfect positive linear relationship.

A significant ($p < 0.05$) decrease in the activities of these enzymes was demonstrated in stressed non-treated rats. Paroxetine-treated group showed that their activities were significantly ($p < 0.05$) increased compared to stressed non-treated group and reached levels comparable to that recorded with control group (1).

This suggests that paroxetine owns a possible antioxidant action in case of chronic restrained albino rats with a significant ($p < 0.05$) decrease in their liver transaminases.

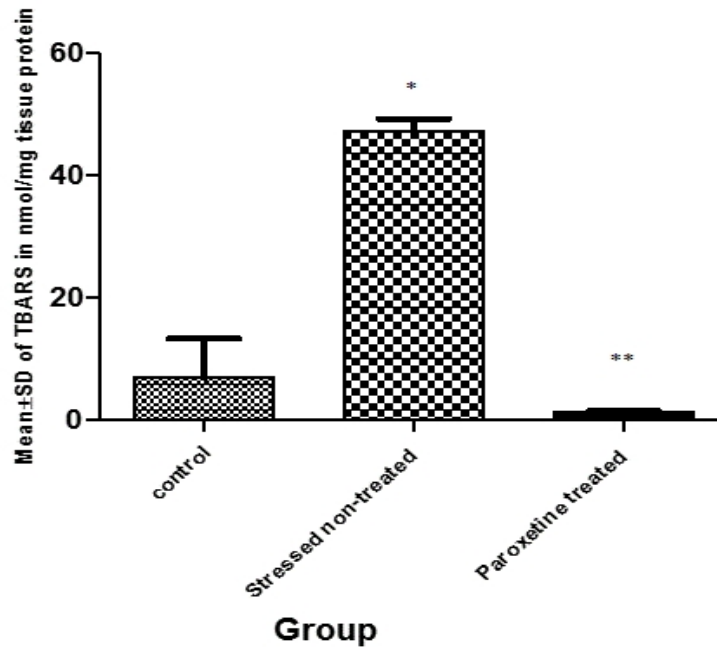


Fig. 2. Effect of 21 days administration of paroxetine on thiobarbituric acid-reactive substance (TBARS) in nmol/mg tissue protein of the liver tissue homogenates of the tested rats.

Fig. 2 showed the results that are expressed as mean \pm SD ($n = 10$ albino rats/group).

A significant ($*p < 0.05$) decrease in TBARS levels of paroxetine-treated group (3) in comparison to the stressed non treated group (2).

- $*p < 0.05$ = significant increase in TBARS levels in group (2) compared to the control non-stressed non-treated rats group (1)
- $**p < 0.05$ = significant decrease in TBARS levels in paroxetine-treated group (3) compared to the stressed non-treated rats group (2)

Pearson correlation = 0.9 with perfect positive linear relationship

Table (2) shows changes in glutathione levels expressed as nmol/ mg protein before and 4 hrs. After exposure to 0.5 mM diethyl maleate (DEM). Data were obtained from hepatic cells isolated from either control, stressed non-treated or stressed + paroxetine-treated group.

Table 2. Effect of treatment with paroxetine on Glutathione repletion ability of hepatic cells expressed nmoles of glutathione per milligram of protein

Groups	Change in glutathione level in hepatic cells		
	Before exposure to DEM	4 hours after exposure to DEM	% change in glutathione level
Control	54.5±2.4	55.4±3.2	+1.65%
Stressed non-treated	53.5±3.4	2.1±0.03*	-96.07%
Stressed paroxetine-treated	51.2±3.1	55.7±3.7**	+8.79%

*There is significant (*p<0.05) reduction in glutathione levels in stressed non-treated rats compared to control group. However, treatment of chronic restrained rats with paroxetine for 21 days significantly (**p< 0.05) is likely to protect hepatic cells against acute depletion of glutathione in comparison to group 2 suggesting that ip paroxetine administration would enhance glutathione replenishing ability in hepatic cells of chronic restrained rats.*

**p<0.05= significant reduction in hepatic glutathione replenishing ability in group (2) compared to the control non-stressed non-treated rats group (1).*

***p<0.05 = significant increase in hepatic glutathione replenishing ability in paroxetine-treated group (3) compared to the stressed non-treated rats group (2).*

pearson correlation = 0.9 with perfect positive linear relationship

4. DISCUSSION

Exposure of albino rats, in the present study, to chronic restraint model significantly decreases serum levels of liver transaminases (ALT &AST) and reduces activities of CAT and GPx anti-oxidant enzymes in hepatic homogenates of these stressed rats. It also decreases hepatic replenishing ability of glutathione. Ip Administration of paroxetine for 21 days reverses all these changes to almost the levels of control non stressed group.

Administration of paroxetine ip for 28 days for rats exposed to 3-weeks CMS, either due to acute dexamethasone administration or due to exposure to chronic mild stress (CMS) model of anhedonia, reversed the anhedonic effect. These results provide an evidence about the antidepressant effect of paroxetine as already well-known. Additionally, this study demonstrates its antidepressant effect against anhedonia-induced by glucocorticoids that could be related to suppression of oxidative stress associated with dexamethasone administration [18].

There is a reported relationship between depression and immune system. Results of previous studies revealed that depressed mood is often associated with an activation of the immune system functions with a marked secretion of inflammatory cytokines [19,20]. This increase in immune functions was thought to be related to damaging actions of exposure to oxidative stress. As it was found that activation of immune system results in stimulation of marked synthesis of ROS, instability of anti-oxidant enzymes and down-regulation of their synthesis [8]. Proper control of the functions of immune system leads to a positive impact on the synthesis and activities of the antioxidant enzyme mitochondrial manganese superoxide dimutase (SOD2) [21].

Chronic paroxetine administration to stressed adult rats would produce its protective effects against apoptosis by its possible action against oxidative stress in their hippocampus [22]. These findings could be partially applied to the results of the present study in relation to the possible hepatoprotective effect of paroxetine in chronic restrained rats by its preservation of

antioxidant enzymes and a significant reduction in hepatic TBARS, as a marker of lipid peroxidation.

Male Sprague-Dawley rats were exposed to chronic unpredicted stress with administration of Paroxetine (1.8 mg/kg once daily) by an intragastric gavage. The study recorded any change in rat behaviors, activities of some anti-oxidant enzymes : superoxide dismutase and catalase and lipid peroxidation in the form of malondialdehyde level in the serum in addition to the expression of serotonin transporter (5-HTT) in the hippocampus and norepinephrine transporter (NET) in the pons. Results demonstrated that paroxetine produced a significant reduction in depression-like behaviors and in the malondialdehyde level. Additionally, it increased the activities of superoxide dismutase and catalase anti-oxidant enzymes with an increase in 5-HTT and NET expression. The study concluded that the antidepressive effect of paroxetine could be related partly to overcoming oxidative stress disorders and increasing the expression of 5-HTT and NET that elevated mood of depressed rats [23].

Clinically, chronic administration of paroxetine would protect against apoptosis, oxidative stress and improve cytoskeletal functions, intracellular signaling and serotonergic and catecholaminergic neurotransmission. Paroxetine preserves antioxidant enzymes and plays a vital role in the protection of cells from oxidative stress and hence, it will protect intracellular components such as DNA, proteins and lipids and will prevent disrupting signaling pathways that render each cell in the body susceptible to apoptotic or necrotic cell death. It would also protect the central nervous system from the damaging effect of oxidative stress that accompany exposure of human being to depressed mood, stressful conditions or systemic diseases as diabetes and its cardiovascular complications [24].

5. CONCLUSION

The results of the present study demonstrated a strong correlation between the positive results of paroxetine on hepatic transaminases, some hepatic anti-oxidant enzymes and its ability to replenish glutathione production in isolated hepatic cells of stressed rats. These results would assume a helpful anti-oxidant protective effect on hepatic tissues of chronic restrained rats. Meanwhile, the results of the present study would be a guide for a clinical trial on depressed patients suffering from hepatic impairment.

CONSENT

Not applicable.

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DISCLOSURE

The author reports no conflicts of interest in this work.

COMPETING INTERESTS

Author has declared that no competing interests exist.

REFERENCES

1. Mant A, Rendle V, Hall A, Mitchell W, Montgomery P, McManus W, Hickie I. Making new choices about antidepressants in Australia: the long view 1975-2002. *Med. J. Aust.* 2004;181:21–24.
2. Fava M, Kendler K. Major depressive disorder. *Neuron* 2000;28:335–341. doi: 10.1016/S0896-6273(00)00112-4
3. Ellis E. Reactive carbonyls and oxidative stress: potential for therapeutic intervention. *Pharmacol. Ther.* 2007, 115, 13–24. doi: 10.1016/j.pharmthera.2007;03:015.
4. Halliwell B. Role of free radicals in the neurodegenerative diseases: therapeutic implications for antioxidant treatment. *Drugs Aging.* 2001;18:685–716. doi: 10.2165/00002512-200118090-00004.
5. Guicciardi M, Deussing J, Miyoshi H, Bronk S, Svingen P, Peters C. Cathepsin B contributes to TNF- α -mediated hepatocyte apoptosis by promoting mitochondrial release of cytochrome c. *J. Clin. Invest.* 2000;106:1127–1137. doi: 10.1172/JCI9914.
6. Gould T, Manji H. Signaling networks in the pathophysiology and treatment of mood disorders. *J. Psychosom. Res.* 2002;53:687–697. doi: 10.1016/S0022-3999(02)00426-9.
7. El Melegy, AAM, Kamal SM. Possible Antioxidant and Glutathione Replenishing Effects of Paroxetine in Liver Homogenates of Rats Exposed to Chronic Restraint Model. *J. Pharma. Res.* 2013;3(1):70-73.
8. Bilici M, Efe H, Koroglu M, Uydu H, Bekaroglu M, Deger O. Antioxidant enzymes and lipid peroxidation in major depression: alterations by antidepressant treatment. *J. Affect. Disord.* 2001;64:43–51. doi: 10.1016/S0165-0327(00)00199-3.
9. Fontella F, Siqueira I, Vasconcellos A, Tabajara A, Netto A, Dalmaz C. Repeated restraint stress induces oxidative damage in rat hippocampus. *Neurochem. Res.* 2005;31:105–111. doi: 10.1007/s11064-004-9691-6
10. Eren I, Naziroglu M, Demirdas A, Celik O, Altunbasak A, Ozmen I, Uz E. Venlafaxine modulates depression-induced oxidative stress in brain and medulla of rat. *Neurochem Res.* 2007;32(3):497-505.
11. Zafir A, Ara A, Banu N. *In vivo* antioxidant status: a putative target of antidepressant action. *Prog. Neuropsychopharmacol. Biol. Psychiat.* 2009;33:220–228. doi: 10.1016/j.pnpbp.
12. David D, Bourin M, Jego G, Przybylski C, Jolliet P, Gardier A. Effects of acute treatment with paroxetine, citalopram and venlafaxine in vivo on noradrenaline and serotonin outflow: a microdialysis study in Swiss mice. *Br. J. Pharmacol.* 2003;140:1128–1136.
13. Rotruck J, Pope A, Ganther H, Swanson A, Hafeman D, Hoekstra W. Biochemical role as a component of Glutathione peroxidase. *Science*, 1973;79:588–590.
14. Sinha KA. Colorimetric assay of catalase. *Ann Biochem.* 1972;47:389–394.

15. Fraga C, Leibovitz B, Tappel A. Lipid peroxidation measured as thiobarbituric acid reactive substances in tissue slices: characterization and comparison with homogenates and microsomes. *Free Radic Biol Med*. 1988;4:155–161.
16. Saville B. A scheme for the colorimetric determination of microgram amounts of thiols., *The Analyst*. 1958;83:670-672.
17. Bradford M. A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Anal Biochem*. 1976;72:248–254.
18. Casarotto PC, Andreatini R. Repeated paroxetine treatment reverses anhedonia induced in rats by chronic mild stress or dexamethasone. *Eur Neuropsychopharmacol*. 2007;17(11):735-742.
19. Smith SM, Vale WW. The role of the hypothalamic-pituitary-adrenal axis in neuroendocrine responses to stress. *Dialogues Clin Neurosci*. 2006;8(4):383-95.
20. Maes M, Scharpè S, Meltzer H, Okayli G, Bosmans E, D'Hondt P. Increased neopterin and interferon- gamma secretion and lower availability of L-tryptophan in major depression: further evidence for an immune response. *Psychiatry Res*. 1994;54:143–160. doi: 10.1016/0165-1781(94)90003-5.
21. Bogunovic B, Stojakovic M, Chen L, Maric, M. An unexpected functional link between lysosome thiol reductase and mitochondrial manganese superoxide dismutase. *J. Biol. Chem*. 2008;283:8855–8862. doi: 10.1074/jbc.M708998200.
22. Karanges E, Kashem M, Sarker R, Ahmed E, Ahmed S, Van Nieuwenhuijzen P, Kemp A, Mc Gregor L. Hippocampal protein expression in differentially affected by chronic paroxetine treatment in adolescents and adult rats: A possible mechanism of “paradoxical” antidepressant responses in young patients. *Front. Pharmacol*; 2013. doi: 10.3389/fphar.00086.
23. Qiu HM, Yang JX, Wu XH, Li N, Liu D, Wang LJ, Qin LJ, Zhou QX. Antidepressive effect of paroxetine in a rat model: upregulating expression of serotonin and norepinephrine transporter. *Neuroreport*. 2013;10;24(10):520-5. doi: 10.1097/WNR.0b013e328362066d.
24. Gerö D, Szoleczky P, Suzuki K, Módis K, Oláh G, Coletta C, Szabo C. Cell-based screening identifies paroxetine as an inhibitor of diabetic endothelial dysfunction. *Diabetes*. 2009;62(3):953-64. doi: 10.2337/db12-0789.

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